

# Recovery of functional components from by-products of fruit, vegetable and fish processing

**FIRM Project No: 06RDTAFRC519**

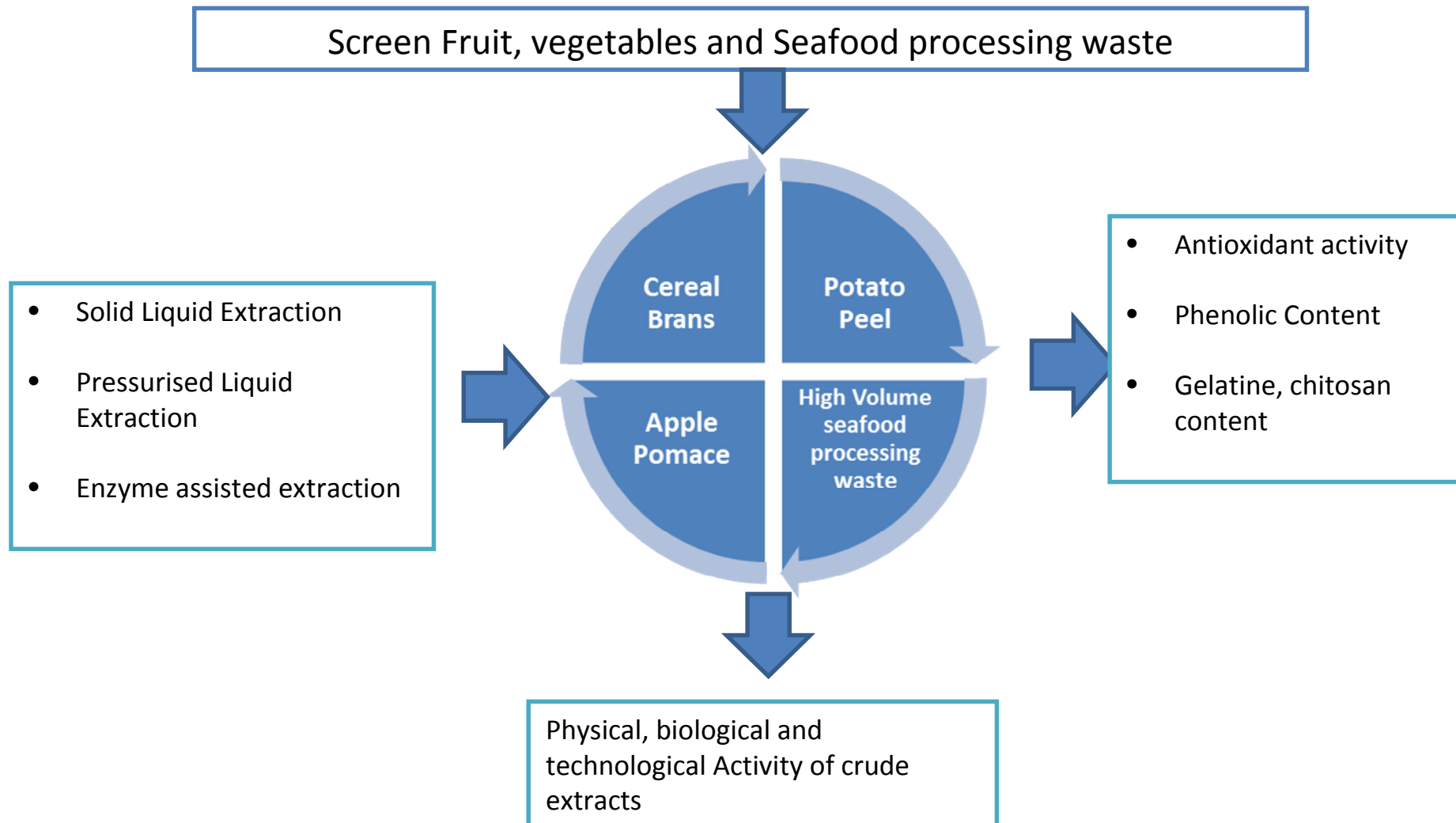
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Ana-Belen Martin-Diana (DIT), Maria Tuohy (NUIG)**



# Outline

- Research approach
- Target sources and compounds
- Screening
- Optimisation of extraction conditions
  - Solid Liquid extraction
  - Pressurised Liquid extraction
  - Enzymatic
  - Extraction of gelatine
- Functional and physical properties of extracted ingredients
- Conclusion

# Research Approach



# Target Sources - Compounds



Potato Peel



Apple Pomace



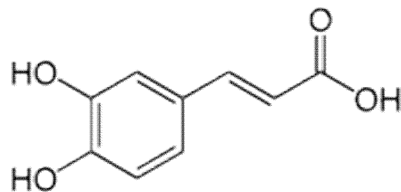
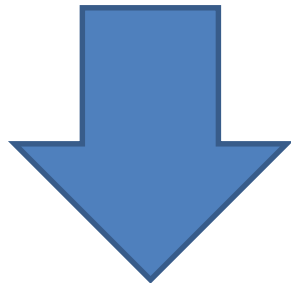
Cereal Brans



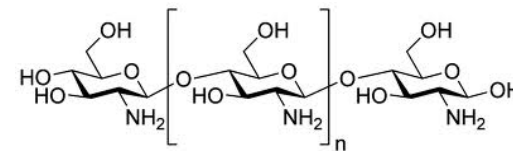
Crab Shell



Pelagic fish processing waste



Poly phenols, antioxidants



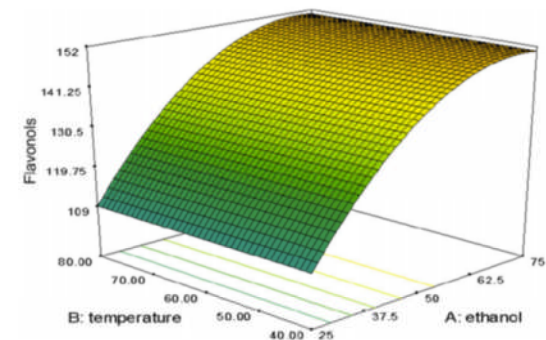
Chitosan



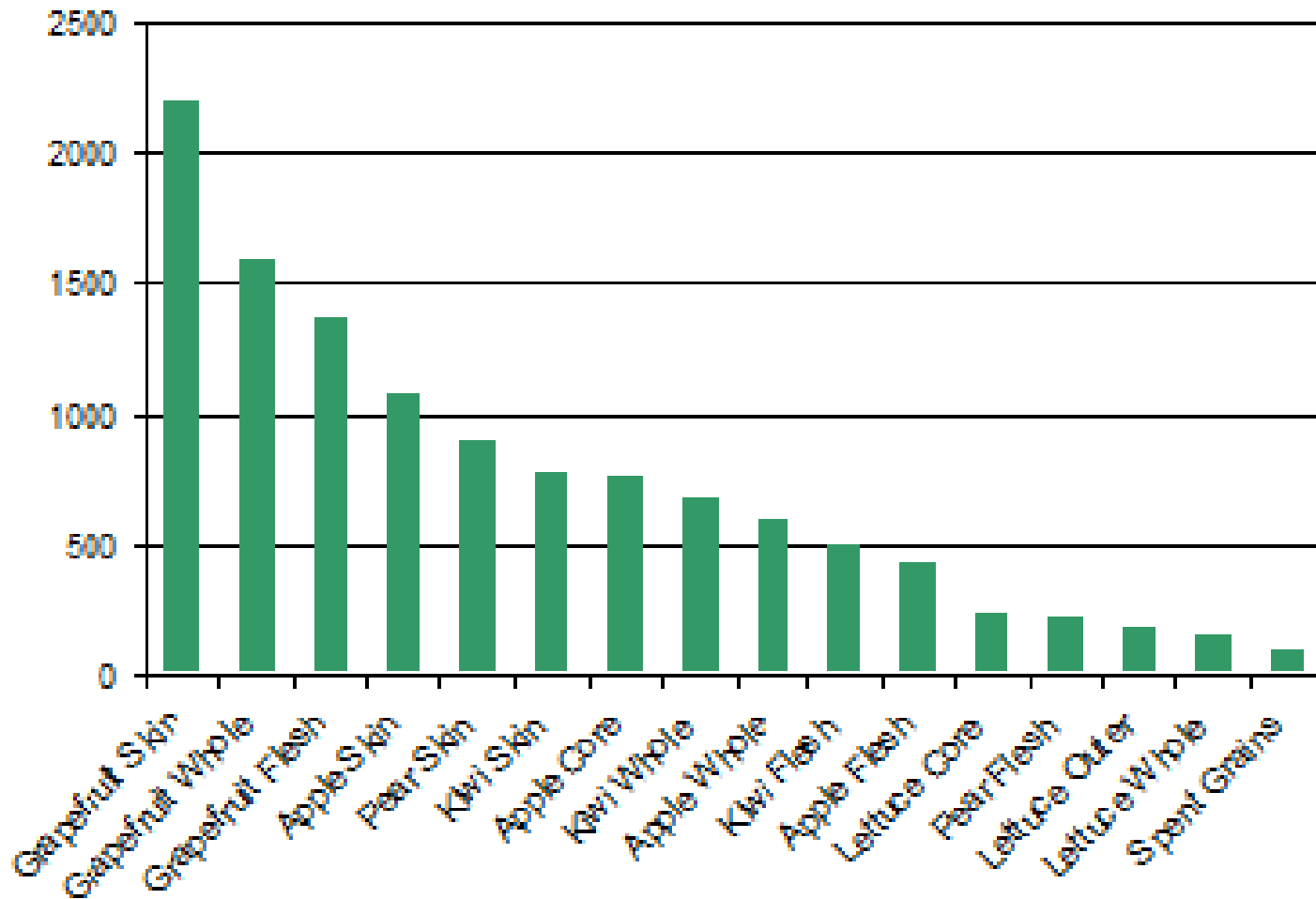
Gelatine

# Response surface methodology

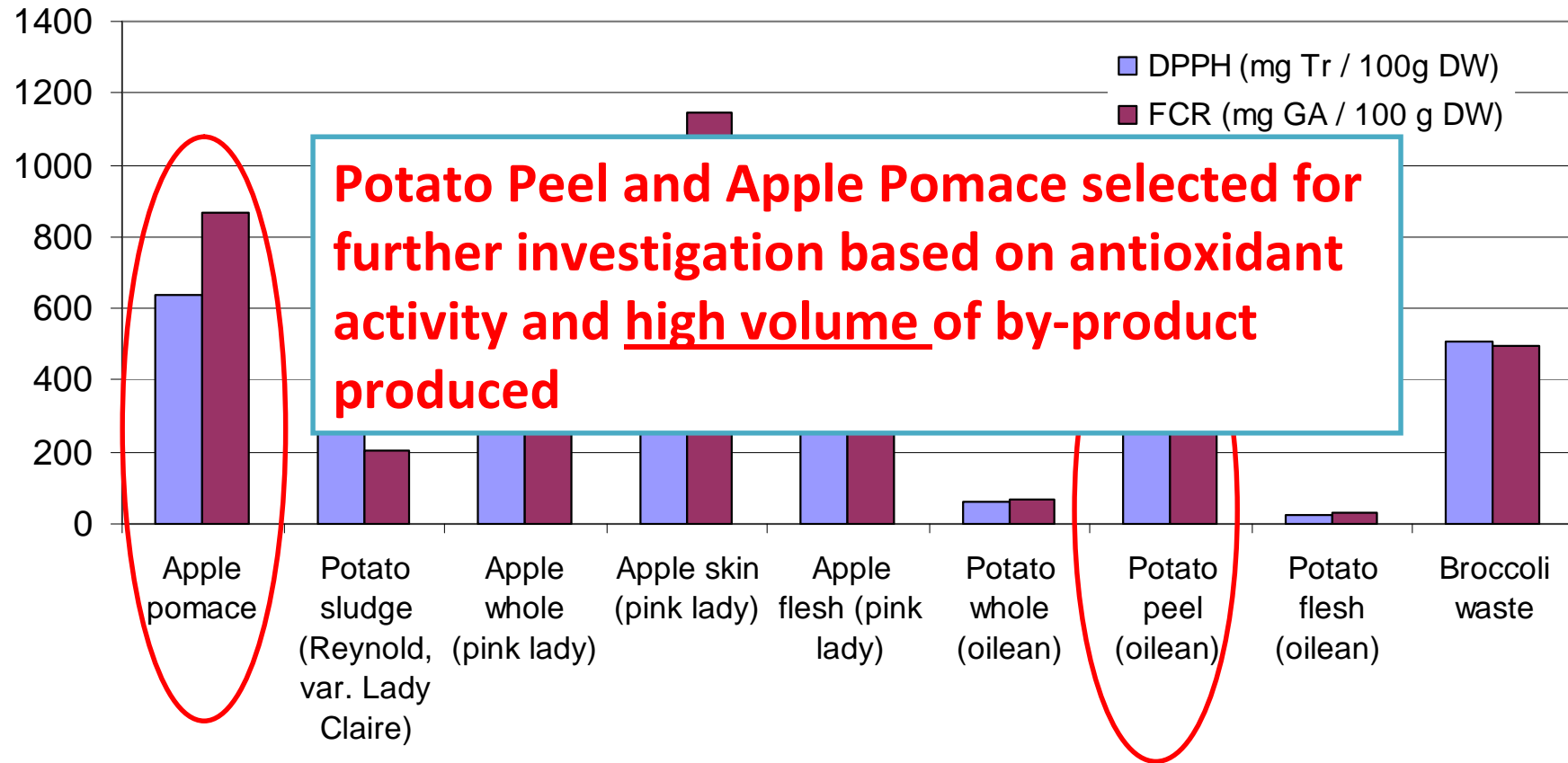
- Statistical technique which uses a series of designed experiments to optimise a response
- Box-Behnken or Central composite design
- No of experiments dependent on number of independent variables
- Results presented a contour plots
- Design Expert 7.1.3
- Determines optimal conditions and predicts the value for a response
- Repeat experiment using optimal conditions to validate model



# Screening – polyphenol content

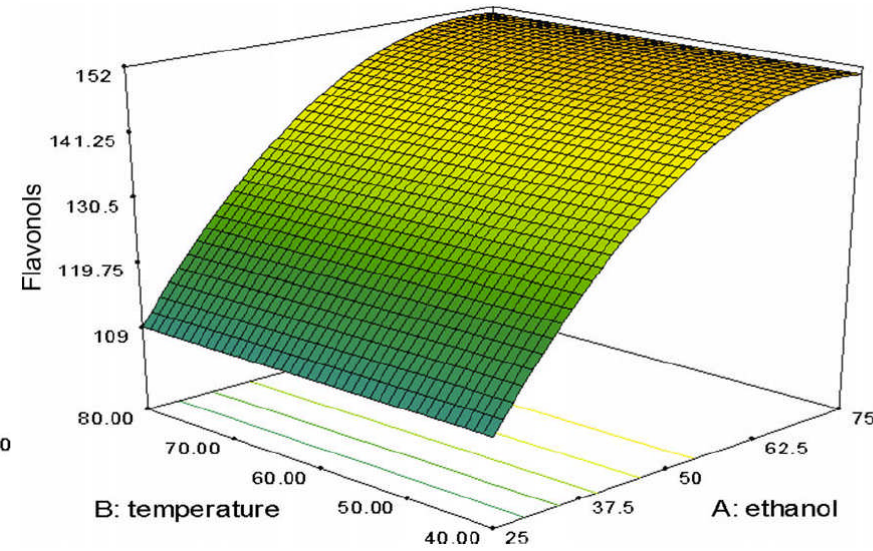
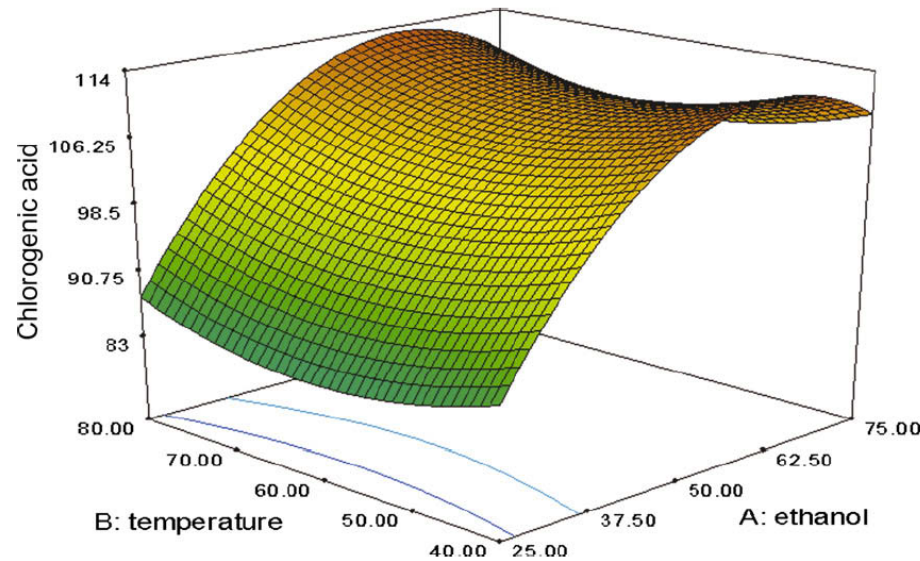


# Screening



Wijngaard, H. H., Rößle, C., & Brunton, N. (2009). *Food Chemistry*, 116(1), 202-207.

# Solid Liquid Extraction – Apple Pomace



**Table 5**

Optimal conditions for extracting compounds with antioxidant activity and the predicted and actual response values with  $X_1$  = ethanol conc. (%),  $X_2$  = temperature ( $^{\circ}$ C);  $X_3$  = time (min); DPPH = antioxidant activity (mg TE/100 g DW); FCR = phenol level (mg GAE/100 g DW); CHA = level of chlorogenic acid ( $\mu$ mol chlorogenic acid/100 g DW); FLA = sum of level of flavonols ( $\mu$ mol rutin/100 g DW); PHLOR = level of phloretin glycoside ( $\mu$ mol phlorizin/100 g DW).

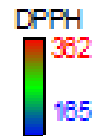
	Response 1 DPPH		Response 2 FCR		Response 3 CHA		Response 4 FLA		Response 5 PHLOR	
	Pred.	Actual	Pred.	Actual	Pred.	Actual	Pred.	Actual	Pred.	Actual
56% Ethanol, 80 $^{\circ}$ C, 27 min	403	449	1065	1092	114	158	147	159	113	100
65% Acetone, 25 $^{\circ}$ C, 60 min	529	436	1511	1415	121	137	177	170	126	124

Wijngaard, H. H. and Brunton, N. 2010. Journal of Food Engineering, 96(1), 134-140



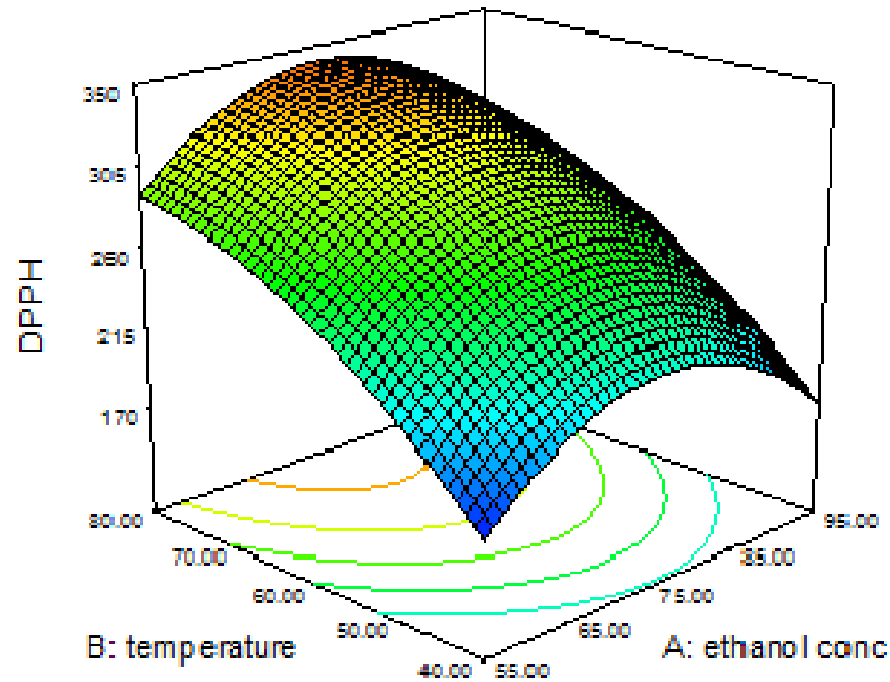
# Solid Liquid Extraction – Potato Peel

Design-Expert® Software



X1 = A: ethanol conc  
X2 = B: temperature

Actual Factor  
C: time = 40.00



**Optimal point for ethanolic extraction was: 75% ethanol, 80°C and 22 min**

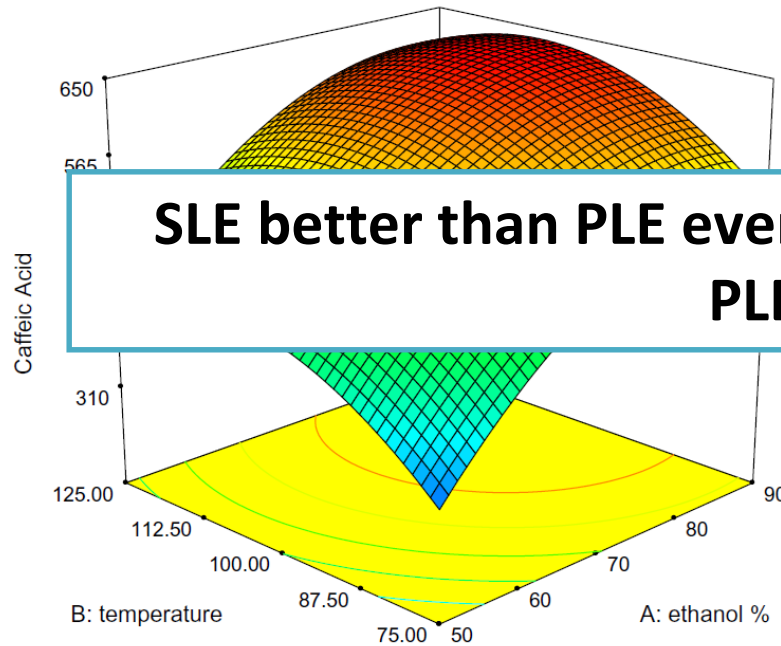
# Pressurised Liquid Extraction

- No solvent evaporation, due to applied pressure
- Higher temperatures can be used than normal
- Better extraction?
- High capital cost
- Upscaling?



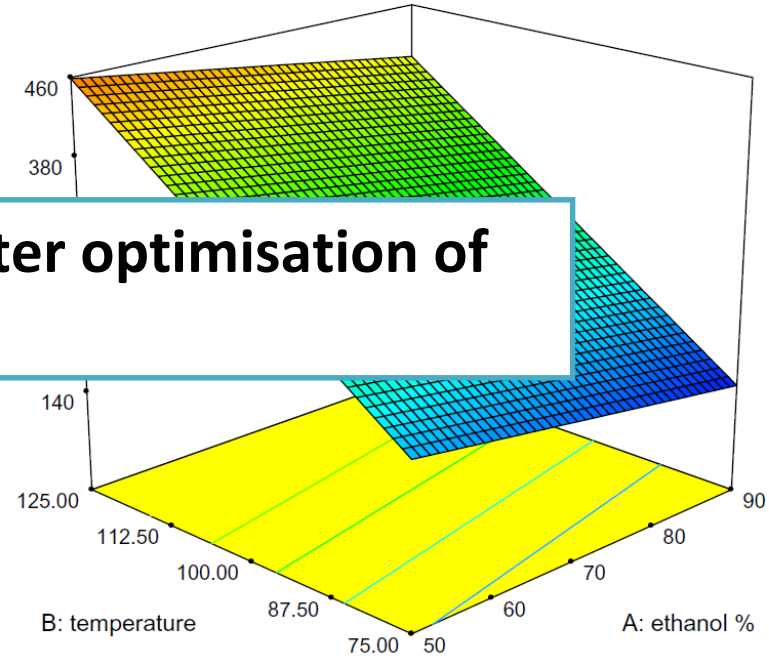
# PLE – Potato Peel

Caffeic Acid



**SLE better than PLE even after optimisation of PLE!**

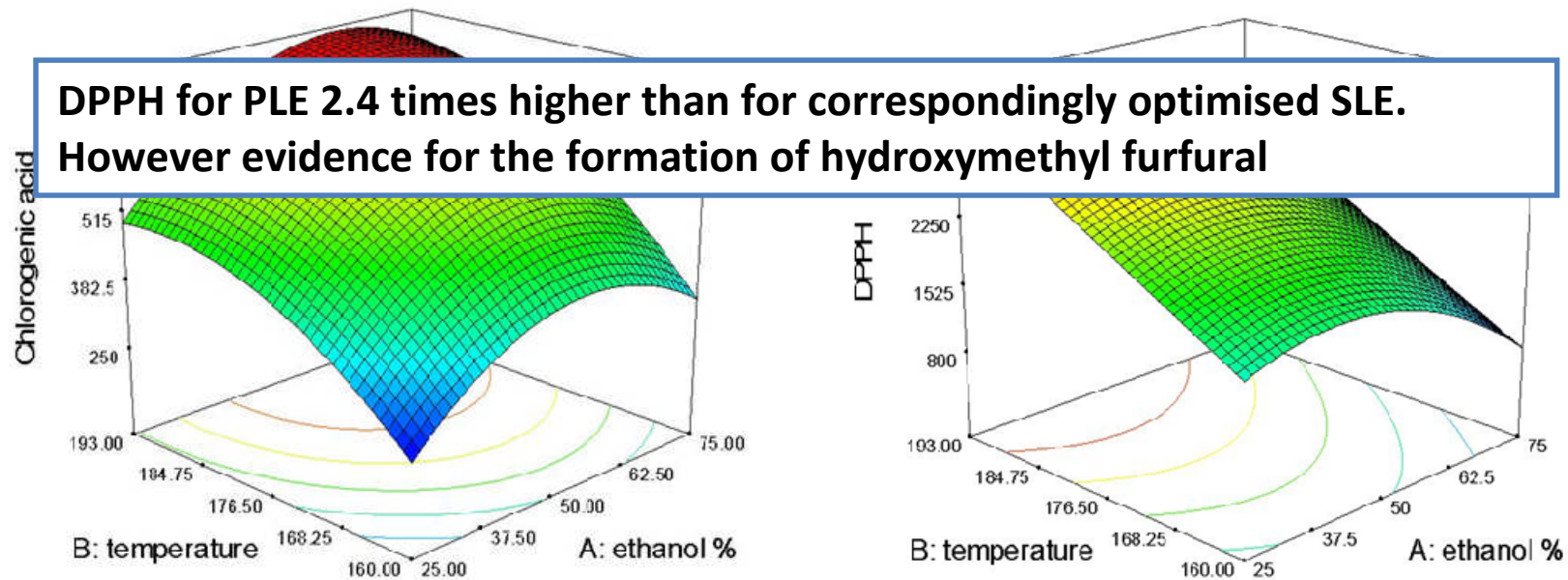
Total Phenolic Content



	Response 1 DPPH		Response 2 FCR		Response 3 CA	
	Pred.	Actual	Pred.	Actual	Pred.	Actual
Conventional extracts: methanolic extracts	NA	200 ± 4	NA	126 ± 7	NA	465 ± 13
Solid-liquid extraction: 75% ethanol, 80 °C, 22 min	355	352 ± 0	409	394 ± 1	606	651 ± 9
PLE: 70% ethanol, 125 °C	365	339 ± 5	431	368 ± 0	573	563 ± 10

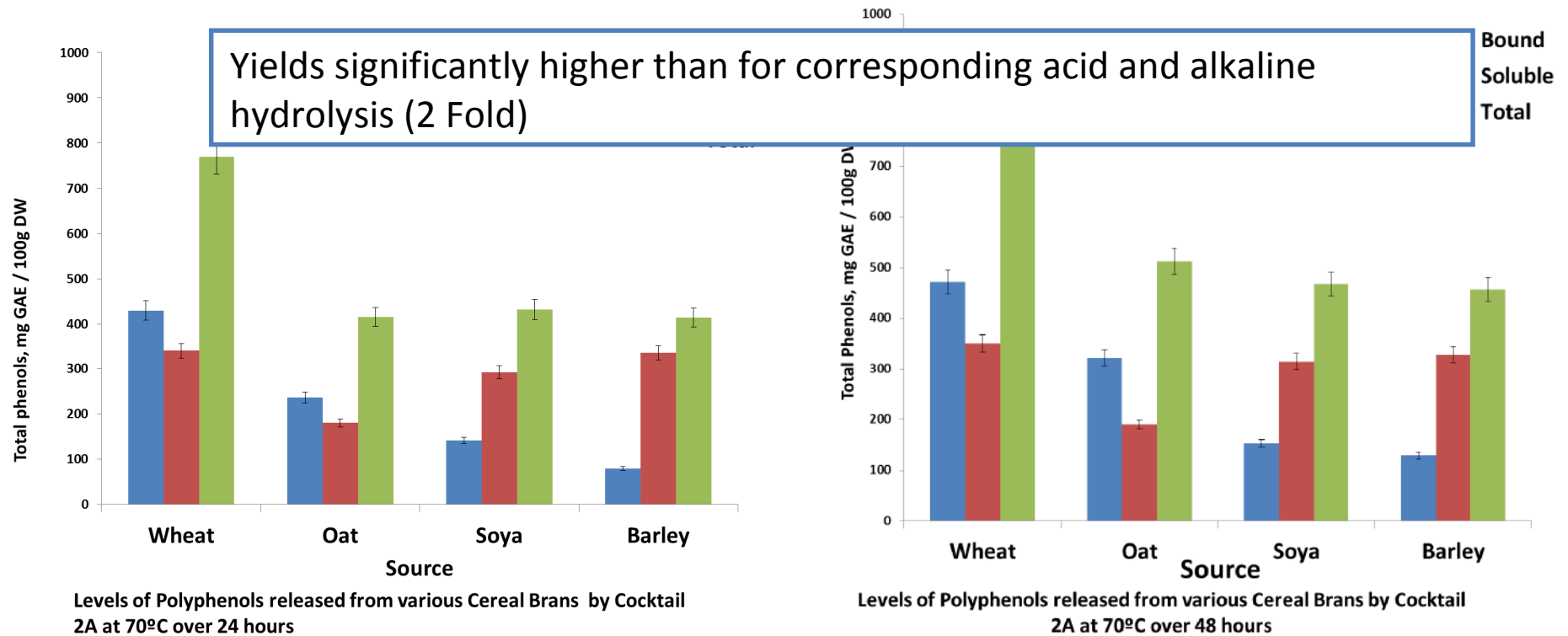
# PLE – Apple Pomace

**DPPH for PLE 2.4 times higher than for correspondingly optimised SLE. However evidence for the formation of hydroxymethyl furfural**

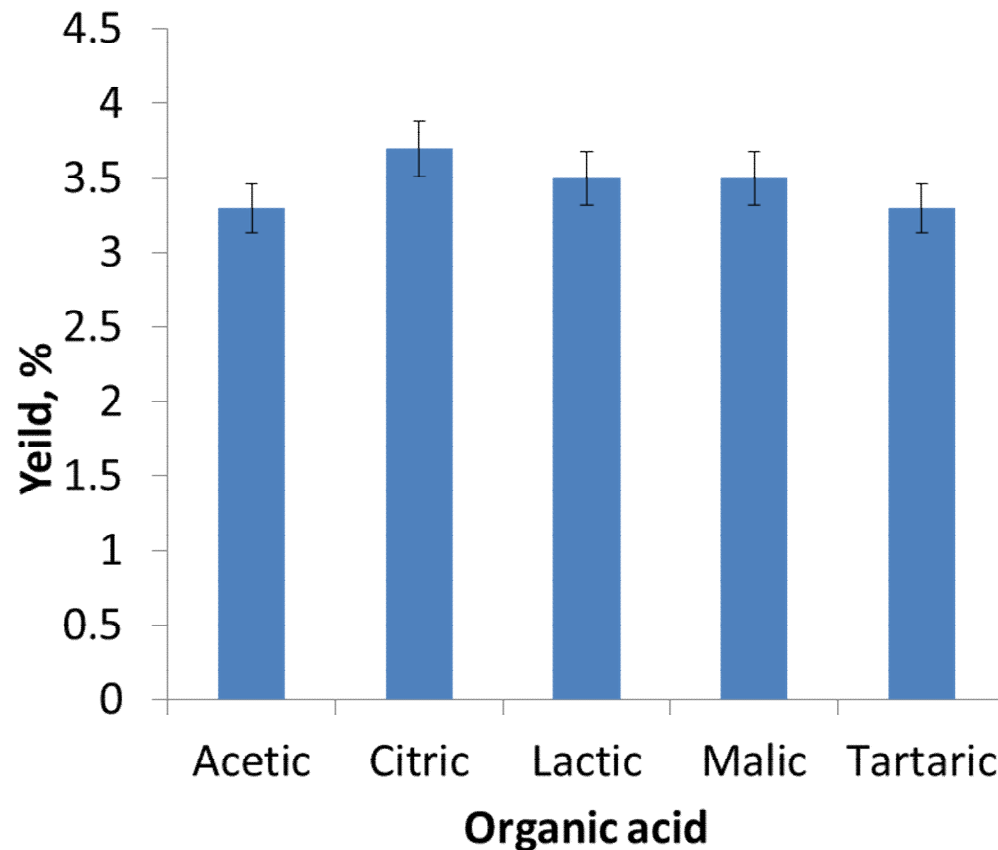


Wijngaard, H., & Brunton, N. (2009). Journal of Agricultural and Food Chemistry, 57(22), 10625-10631.

# Enzymatic extraction – Cereal Brans



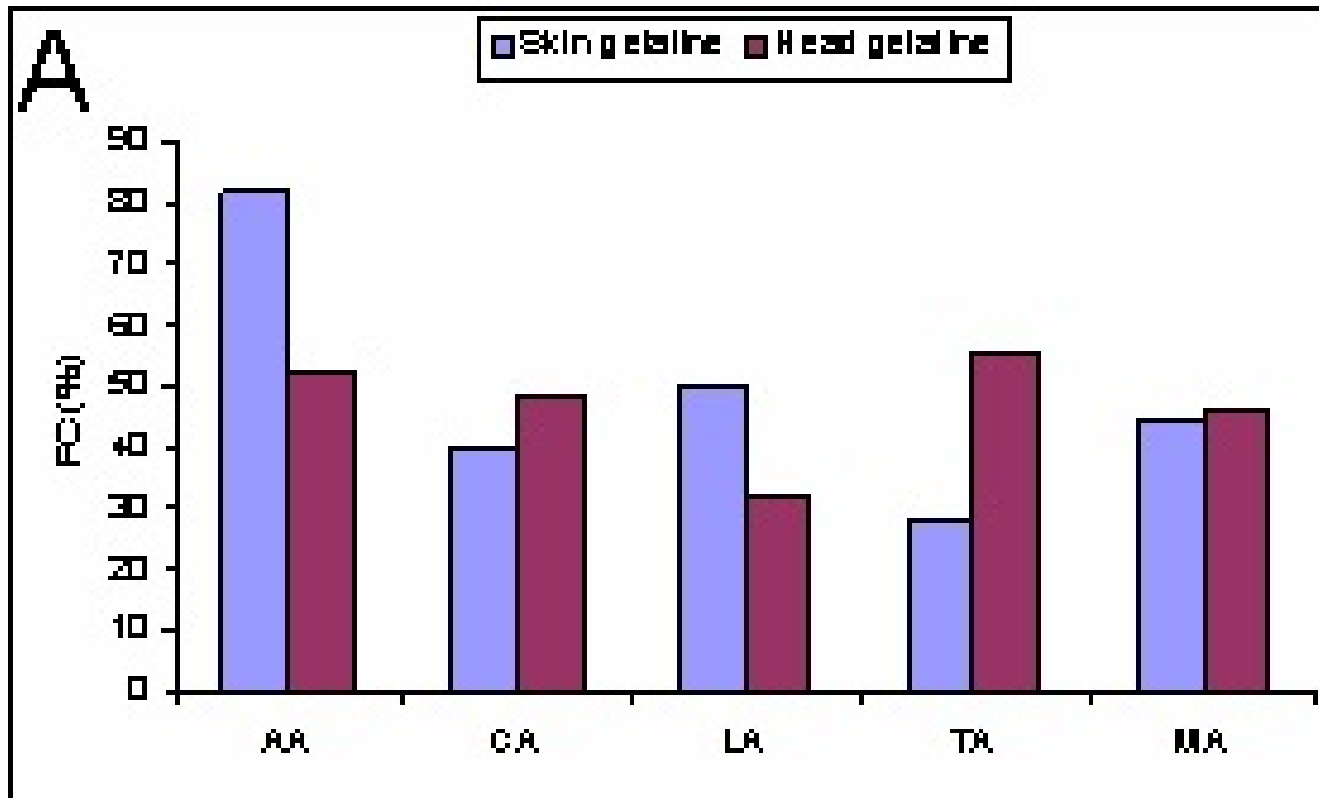
# Extraction of gelatine from mackerel heads



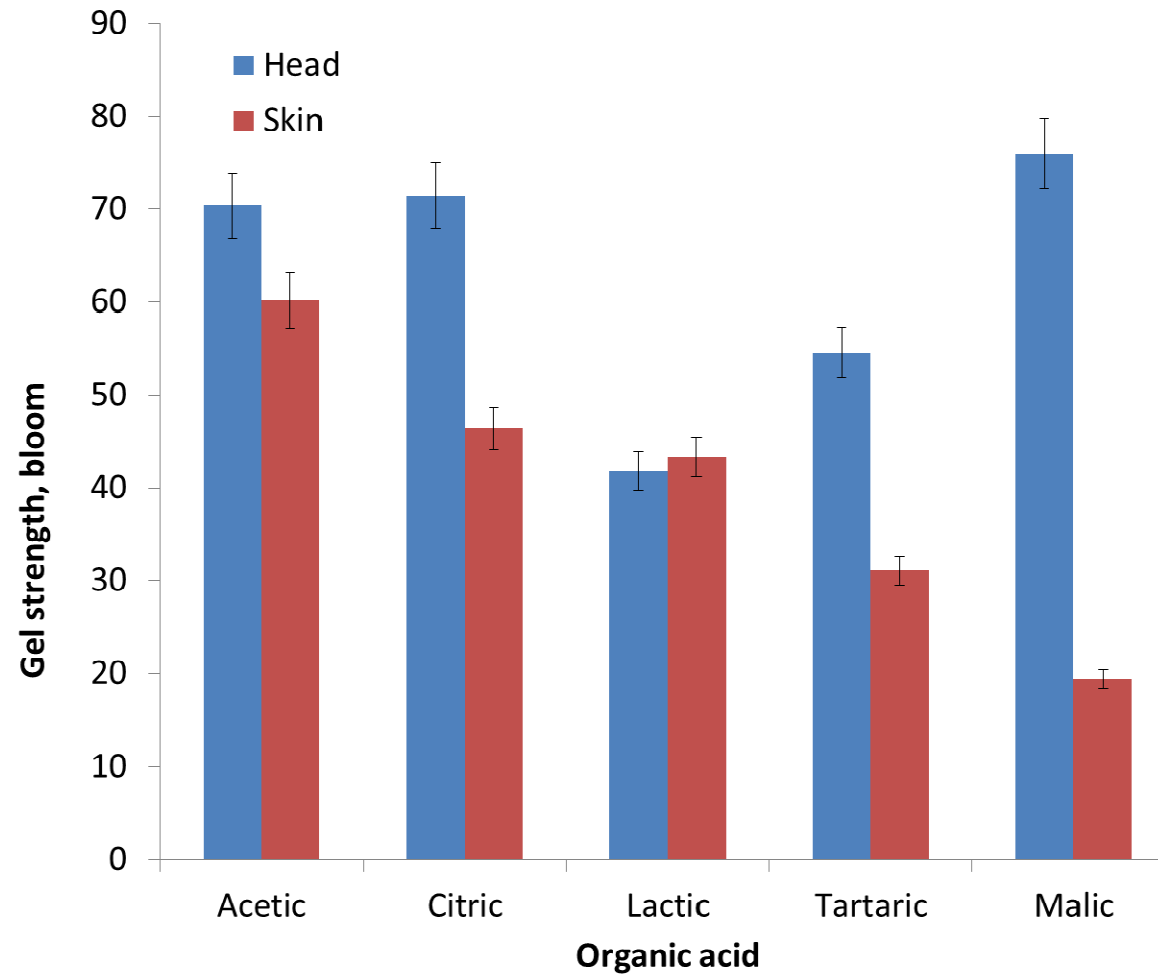
- Lactic and tartaric acids degraded the molecular structure of the gelatines
- Acetic acid showed a significantly ( $p < 0.05$ ) higher  $b^*$  value
- Lactic acid higher turbidity ( $176 \pm 3.2$  FTU),
- Best quality citric and malic acids

Khiari, Zied, et al. Journal of Fisheries Sciences. com 5.1 (2011): 52-63.

# Physical Properties – Extracted Gelatine

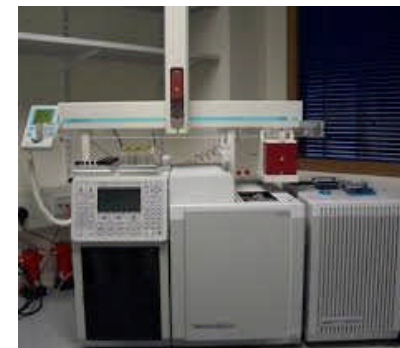
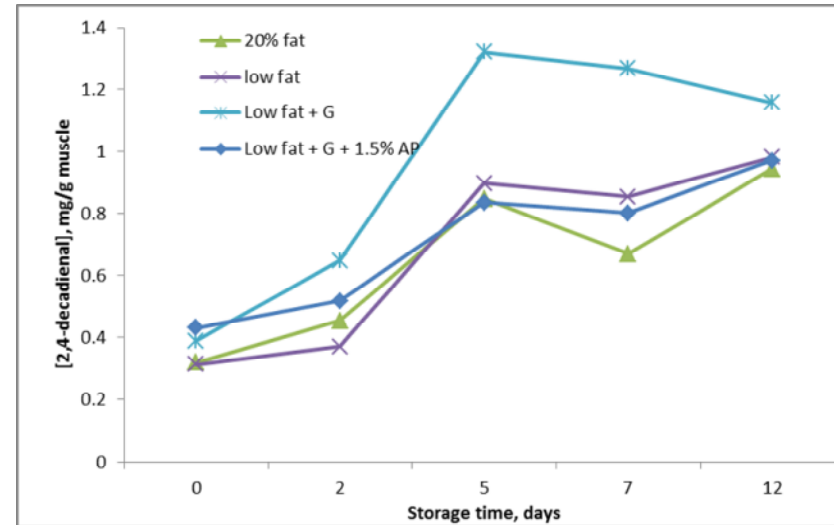
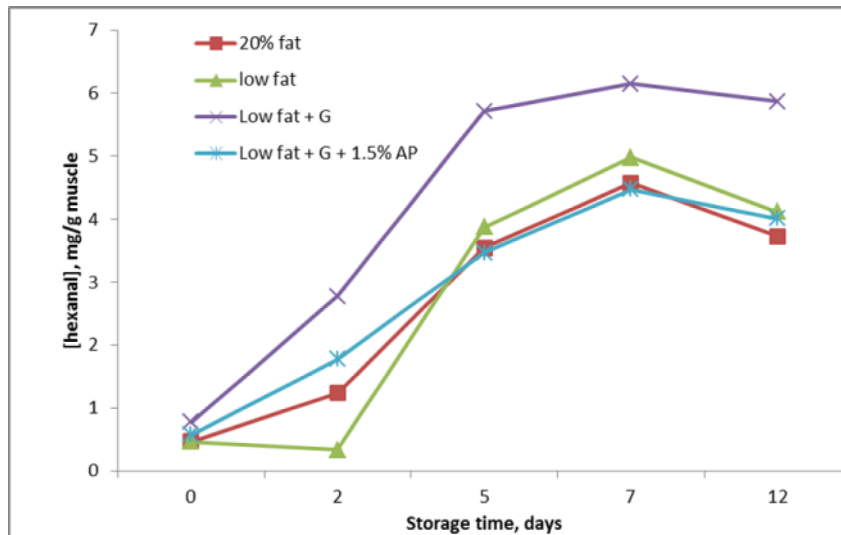
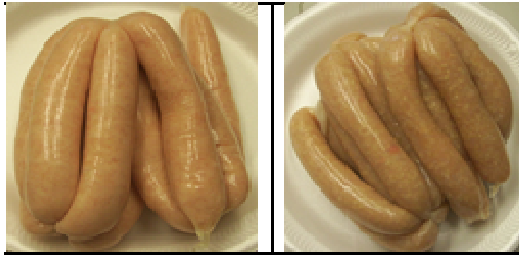


# Physical Properties – Gel strength

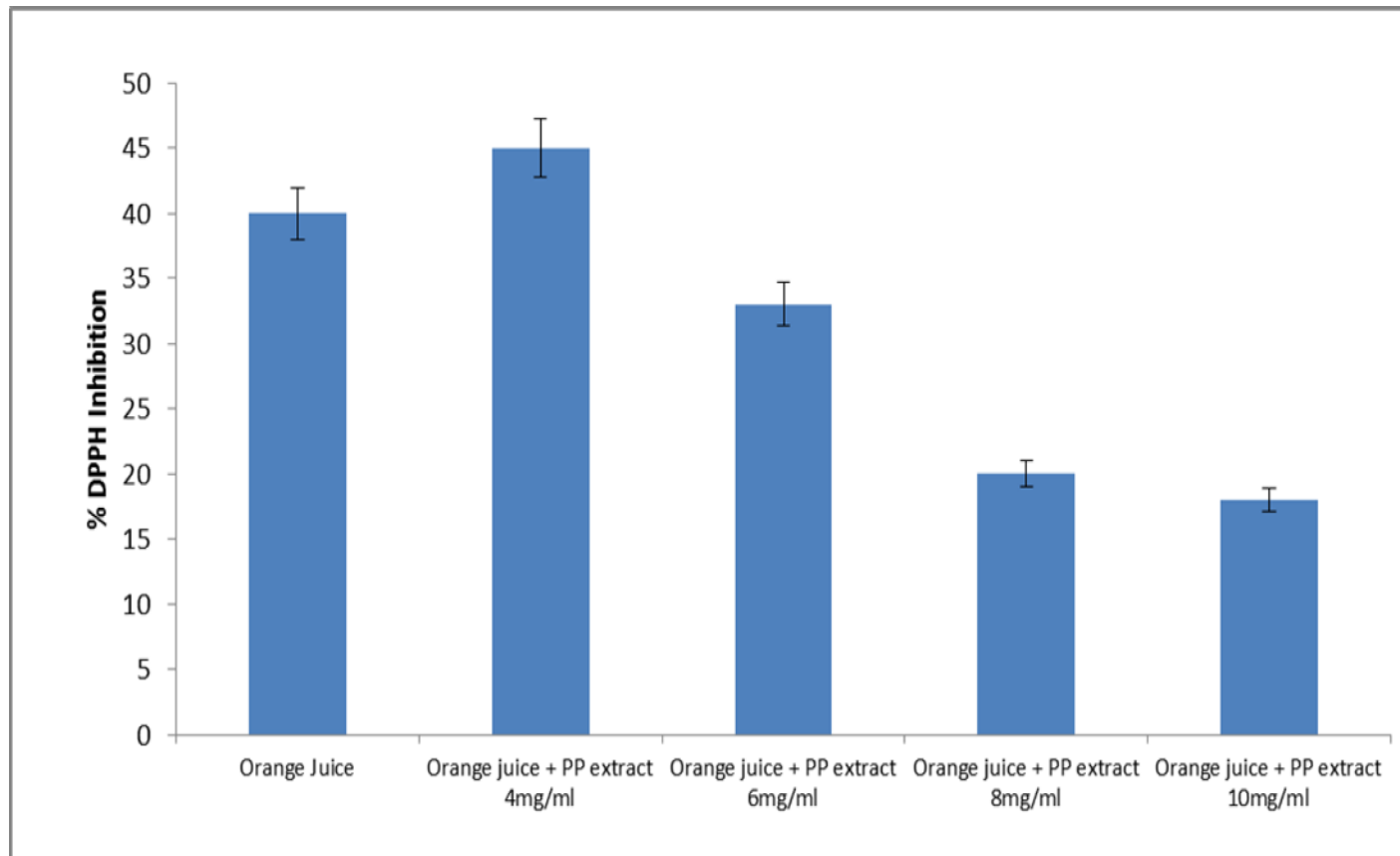




# Functional Properties – Apple Pomace



# Functional Properties – Potato Peel



## Conclusions

- High volume uniform waste required
- Extraction methods – depend on capital cost
- PLE gave better yields for AA's from apple pomace in some cases but resulted in formation of HMF
- Glycoalkoids co-extracted from potato peel
- Supercritical fluid extraction gave low yields for carotenoids from apple pomace

## Conclusions

- Enzyme assisted extraction releases cell wall bound polyphenols from cereal waste
- Yields and functional properties of gelatines extracted from mackerel waste not affected by organic acid but affected by source – head
- AA capacity and lipid stability of foods with extracts from by-products increased.

# Acknowledgements

The Funding for the project was provided under the National Development Plan, through the Food Institutional Research Measure, administered by the Department of Agriculture, Fisheries & Food

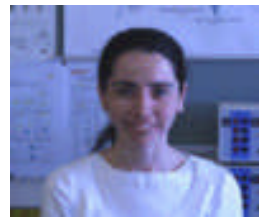
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# Publications

1. Wijngaard, H. H., Rössle, C. and Brunton, N. 2009. A survey of Irish fruit and vegetable waste and by-products as a source of polyphenolic antioxidants. *Food Chemistry*, 116(1), 202-207.
2. Wijngaard, H. and Brunton, N. 2009. The Optimization of Extraction of Antioxidants from Apple Pomace by Pressurized Liquids. *Journal of Agricultural and Food Chemistry*, 57(22), 10625-10631.
3. Wijngaard, H. H. and Brunton, N. 2010. The optimisation of solid-liquid extraction of antioxidants from apple pomace by response surface methodology. *Journal of Food Engineering*, 96(1), 134-140.
4. Khiari, Zied; Rico, Daniel; Martin-Diana, Ana Belen; Barry-Ryan, Catherine. 2010. The extraction of gelatine from Mackerel (*Scomber scombrus*) heads with the use of different organic acids. *Journal of Fisheries Sciences* 5(1): 52-63 (2011).
5. Wijngaard, Hilde Henny, Mélanie Ballay, and Nigel Brunton. (2012). The optimisation of extraction of antioxidants from potato peel by pressurised liquids. *Food Chemistry* 133.4 1123-1130.
6. Wijngaard, H., Hossain, M. B., Rai, D. K., & Brunton, N. (2012). Techniques to extract bioactive compounds from food by-products of plant origin. *Food Research International*, 46(2), 505-513.
7. Khiari, Z., Rico, D., Martin-Diana, A. B., & Barry-Ryan, C. 2013. Comparison between gelatines extracted from mackerel and blue whiting bones after different pre-treatments. *Food Chemistry*, 139 (2013) 347–354