

# Prenatal factors influencing muscle growth & development

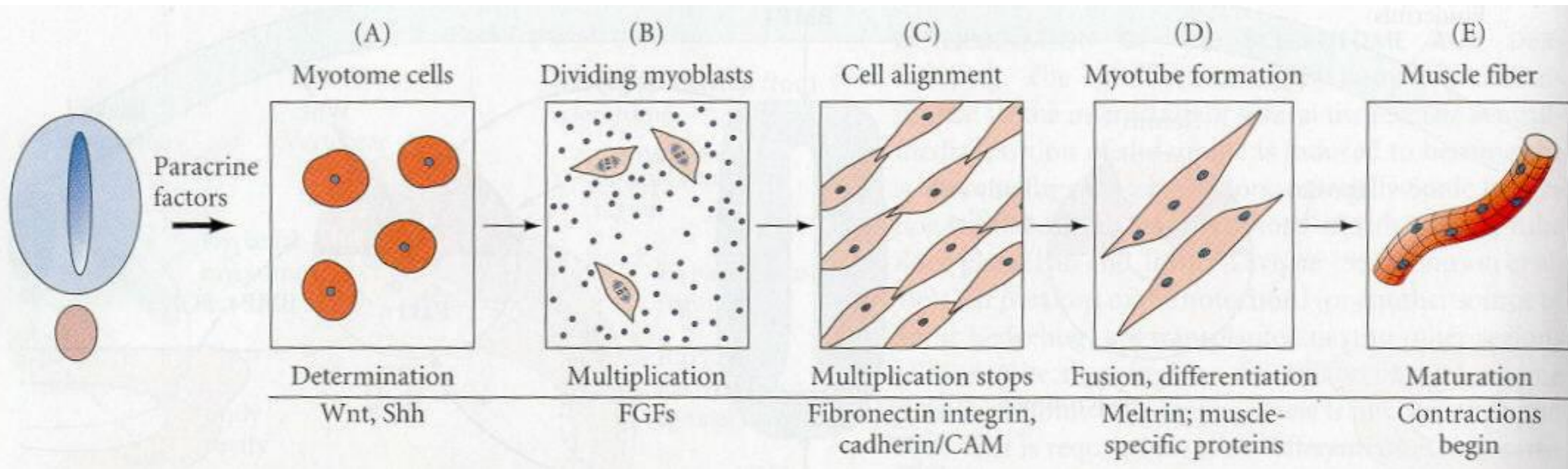
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& Tim Parr

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School of Biosciences

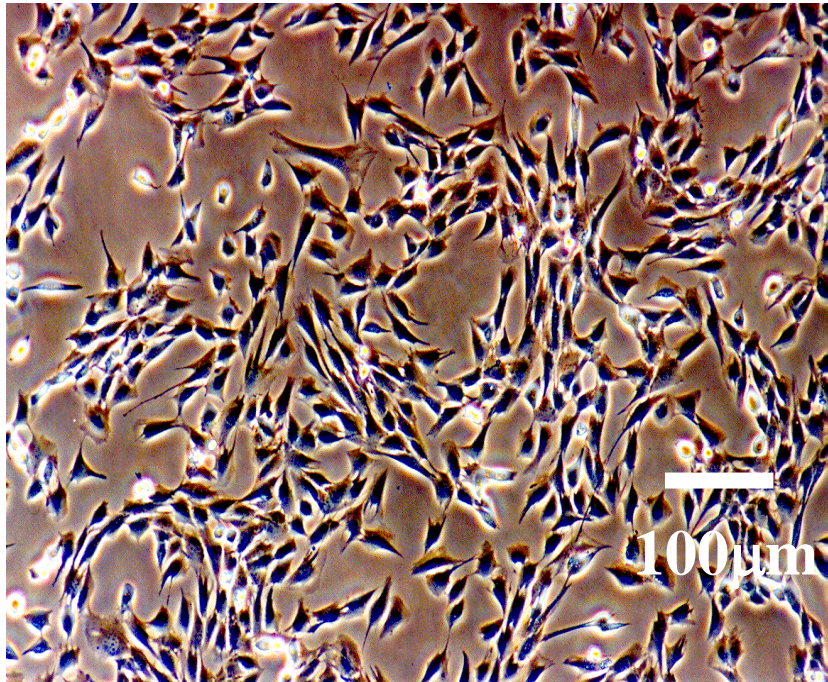
# Outline

- Introduction: Myogenesis & muscle fibre development
- Genetic effects on Muscle Fibre Number
- Maternal Nutrition effects on Muscle Fibre Number
- Long-term effects on muscle wts and adiposity
- Is muscle IGF-II expression a key mechanism?

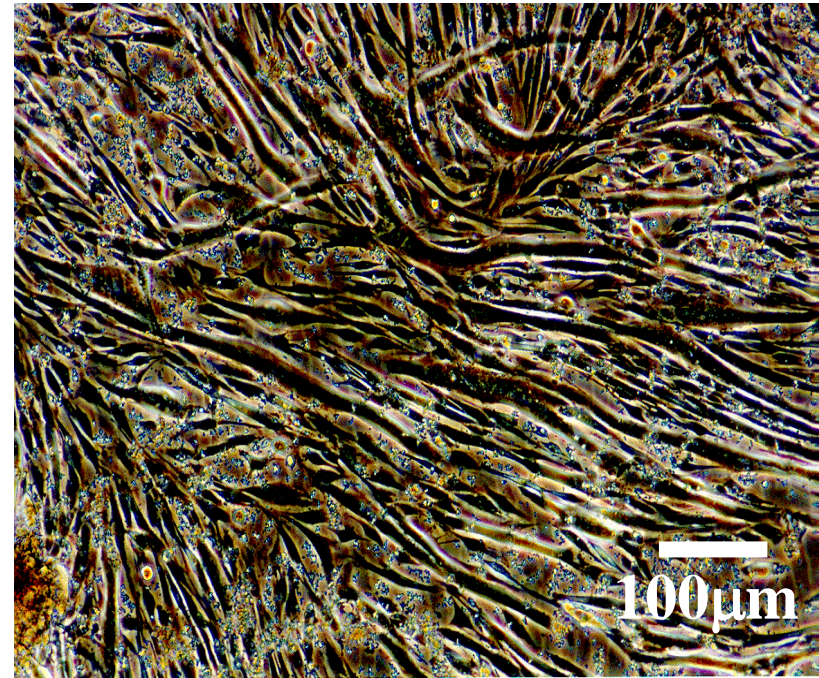
# MYOGENESIS



# Skeletal muscle cells in vitro

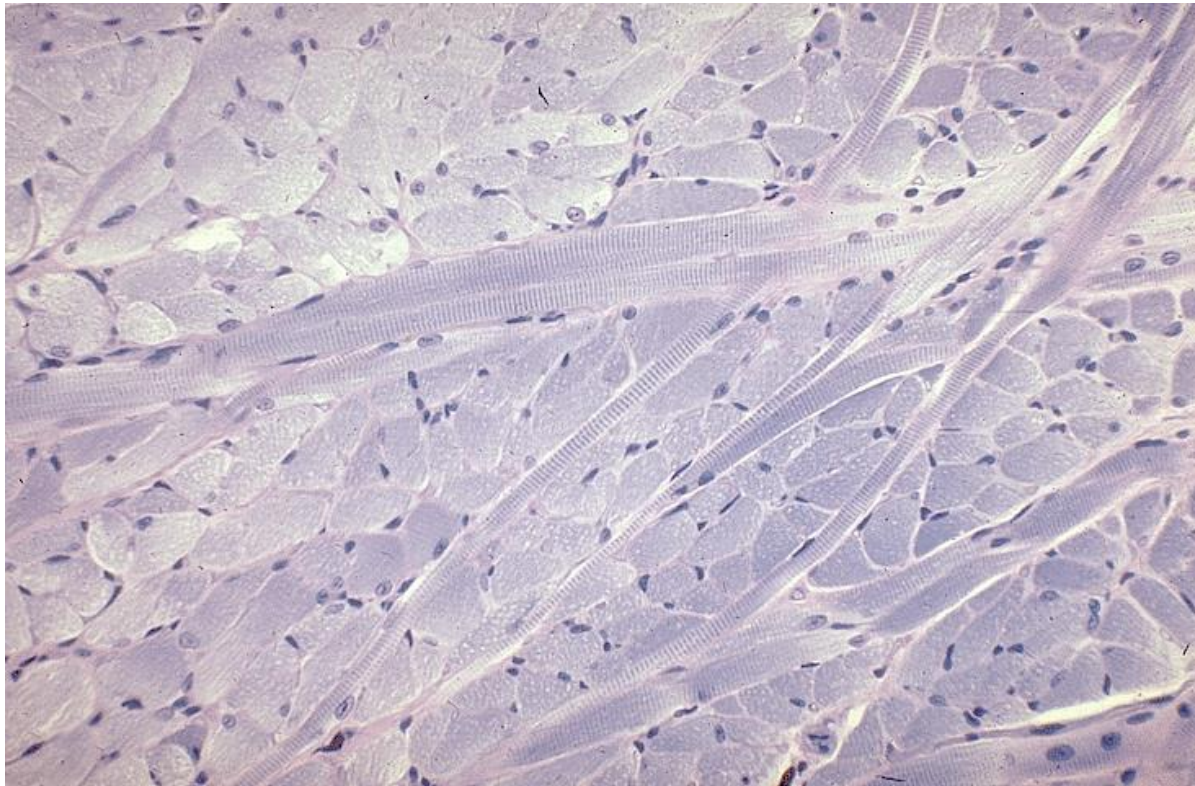


**Proliferating  
(undifferentiated)  
myoblasts**

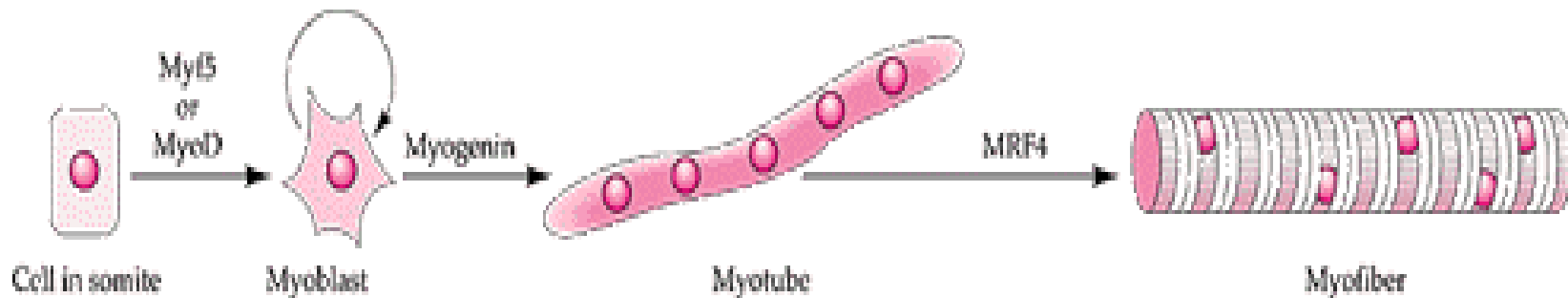


**Differentiated  
myotubes**

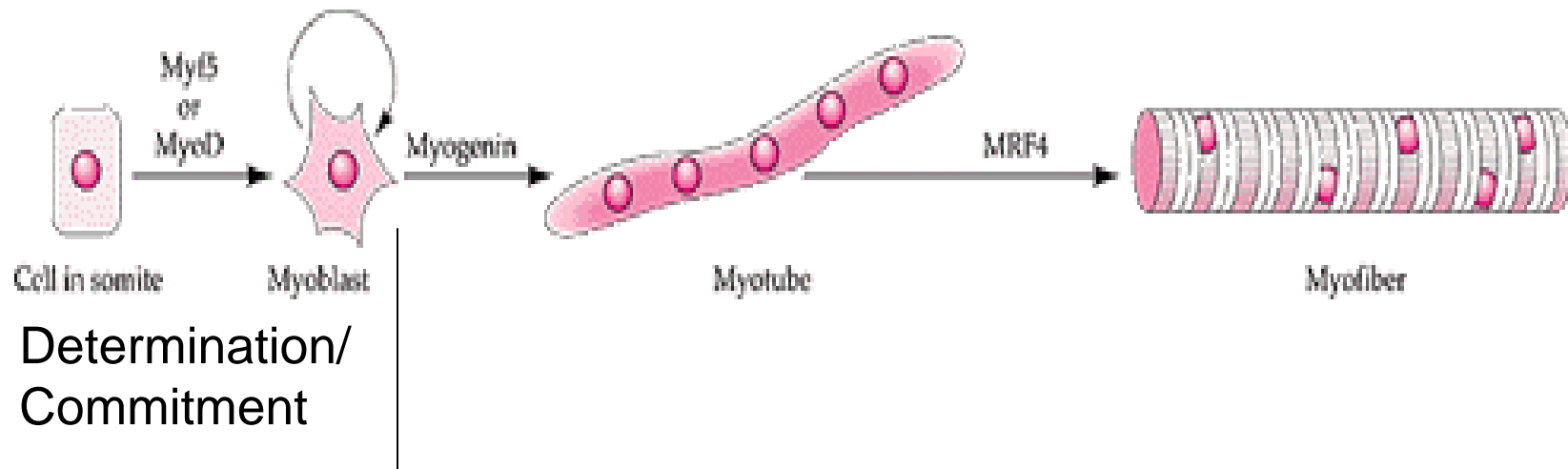
# Section of skeletal muscle



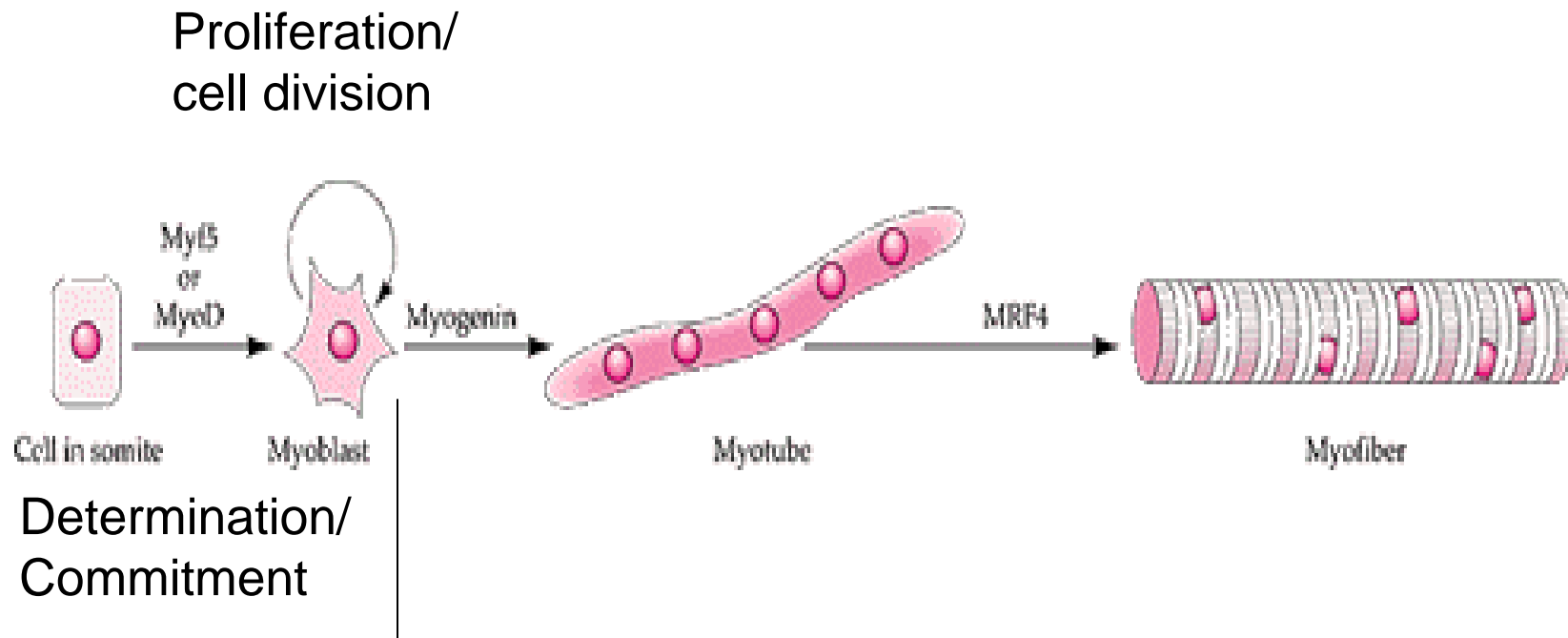
# Regulation of myogenesis



# Regulation of myogenesis

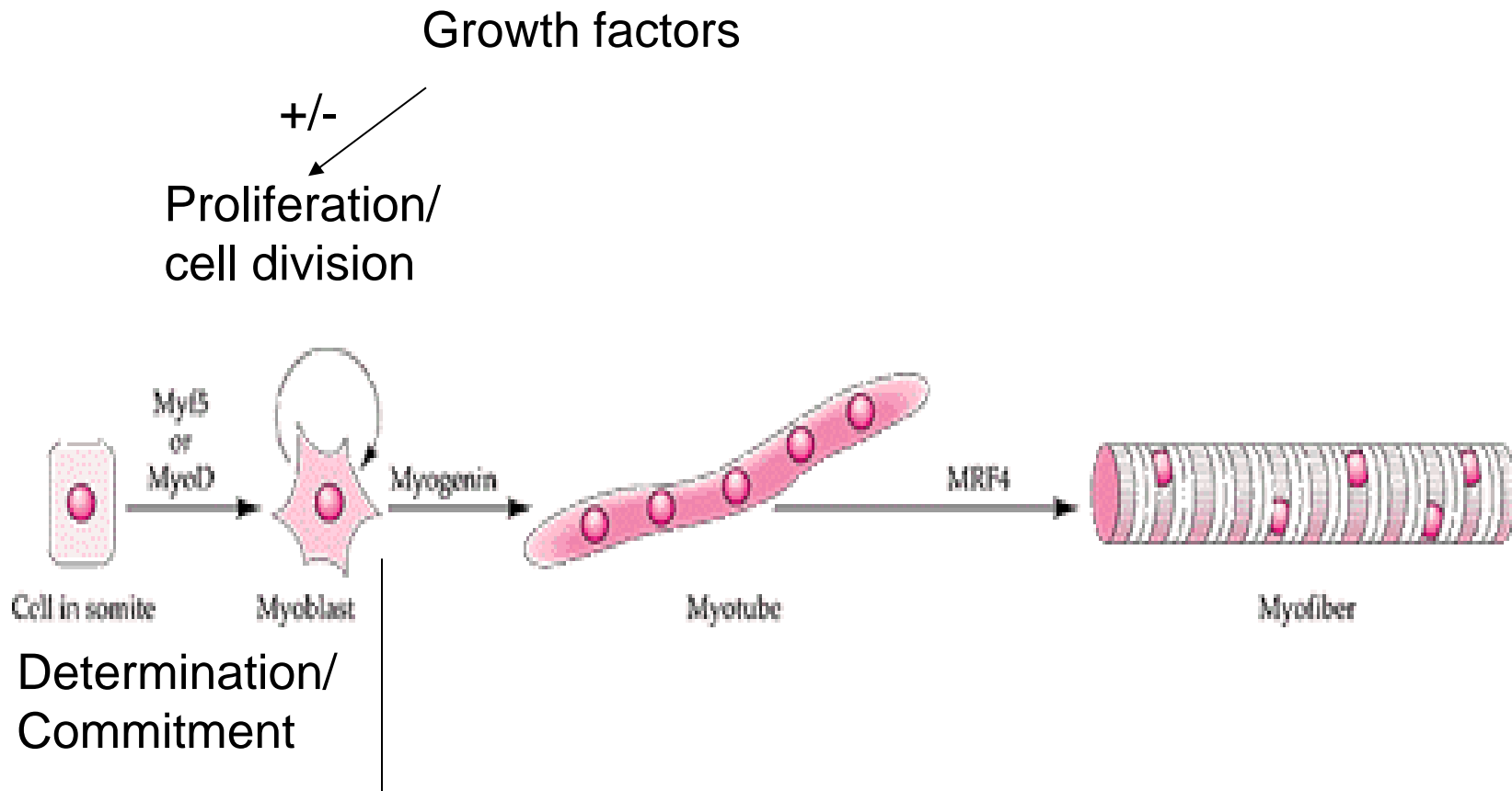


# Regulation of myogenesis

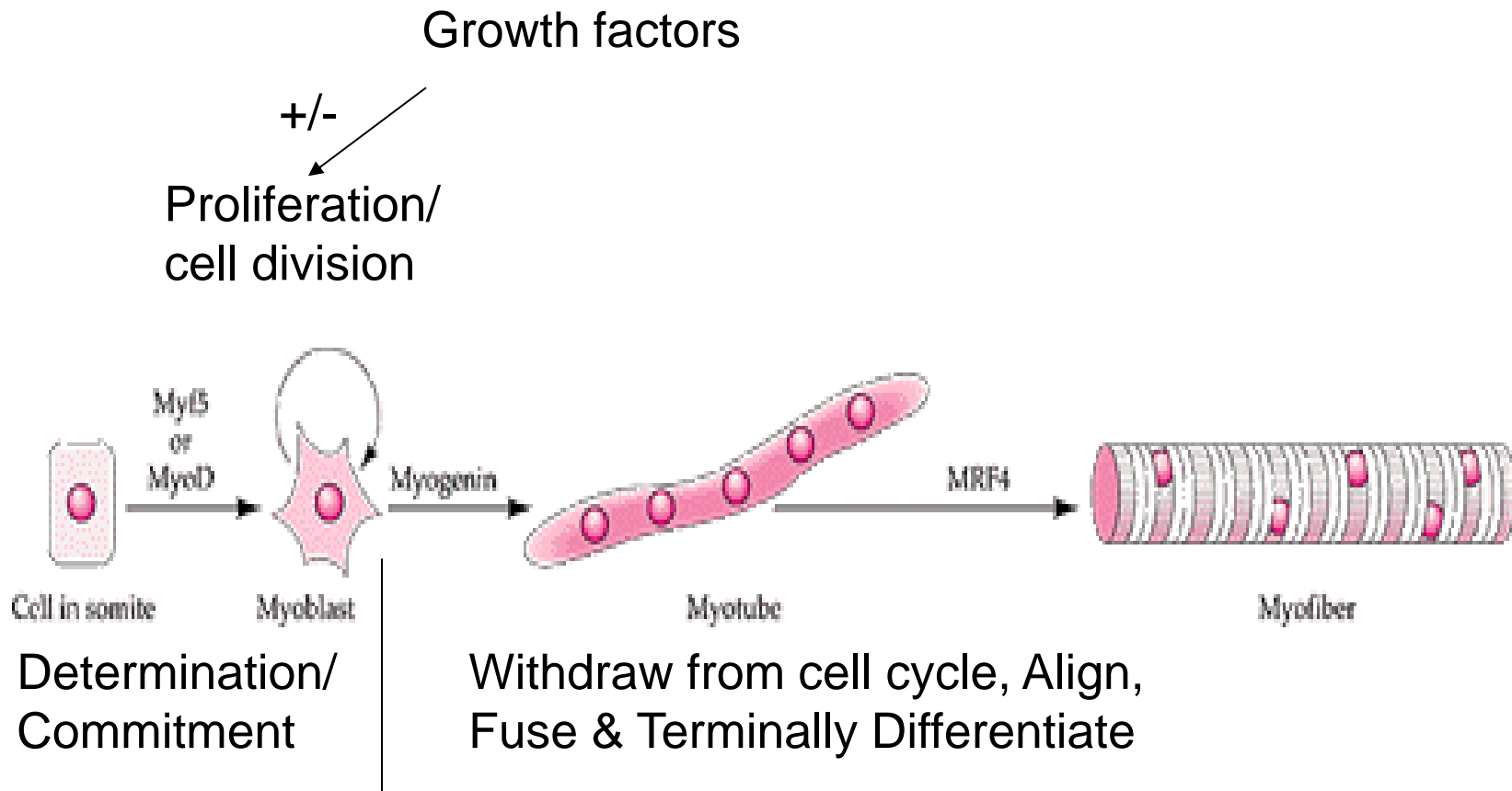




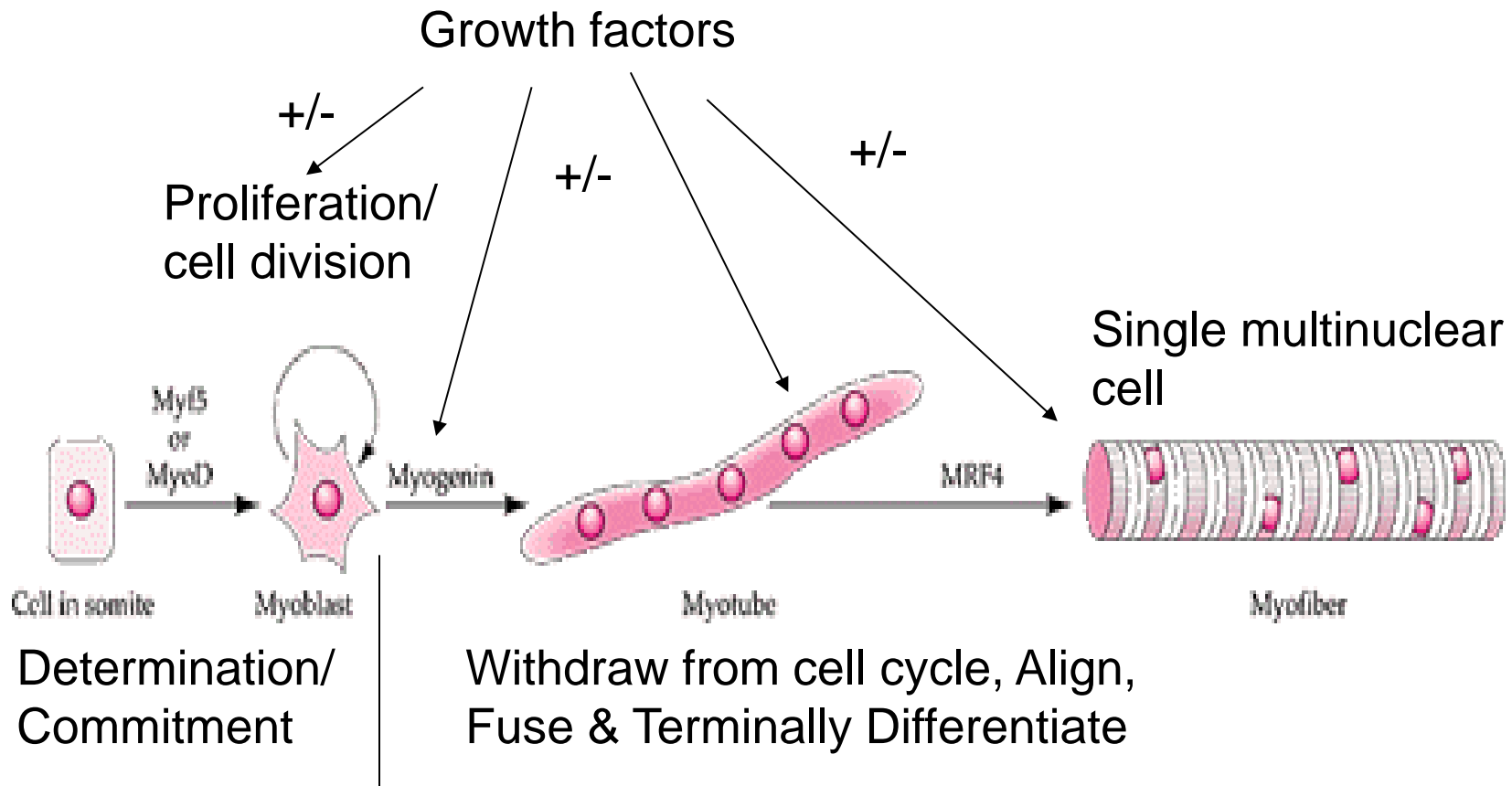
# Regulation of myogenesis



# Regulation of myogenesis



# Regulation of myogenesis



# Growth Factor effects on muscle cell proliferation and differentiation

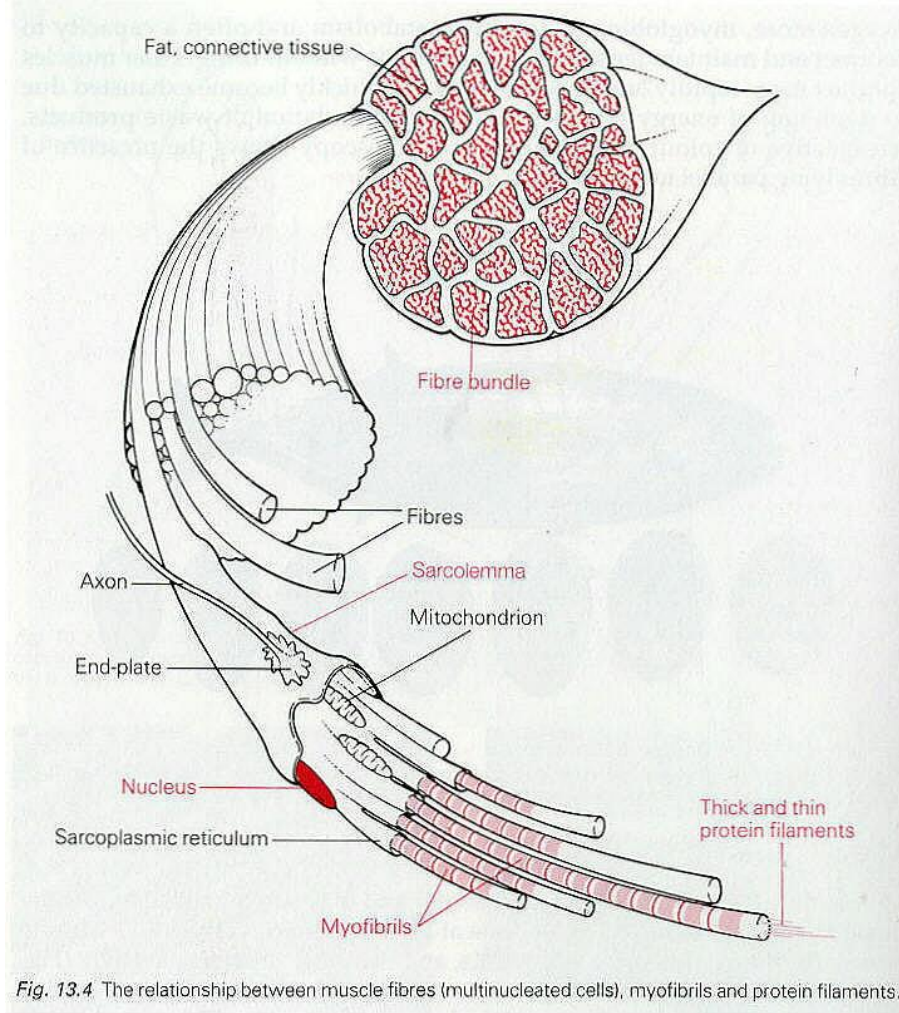
Factor	Precursor cell (myoblast or satellite cell)	
	Proliferation	Differentiation
Insulin	↑	↑
IGF	↑	↑
GH	↑	↑
FGF	↑	↓
EGF and TGF $\alpha$	↑	↑
TGF $\beta$	↓	↓
PDGF	↑	↓
Dex	↑	↑
T3	→	↑
Test	→	↓
$\beta$ -adrenergic agonist	↑	↑
LIF	↑	nd
IL-6	↑	nd
RA	nd	↑
LA	nd	↑
CGRP	nd	↑
TNF- $\alpha$	nd	nd
PGE $_2$	nd	nd
PGF $_{2\alpha}$	nd	nd

## IGF's and myogenesis

- Insulin-like growth factors appear to be unique in that they can act as mitogens and differentiation factors (Florini et al, 1986)
  - concentration-dependent (in cultured cells)
- IGF's shown to increase myogenin expression (cultured cells - Florini et al, 1991a)
- Knockout of IGF-II (transfected with antisense) in spontaneously differentiating muscle cell lines prevents differentiation (autocrine IGF-II effect - Florini et al 1991b)

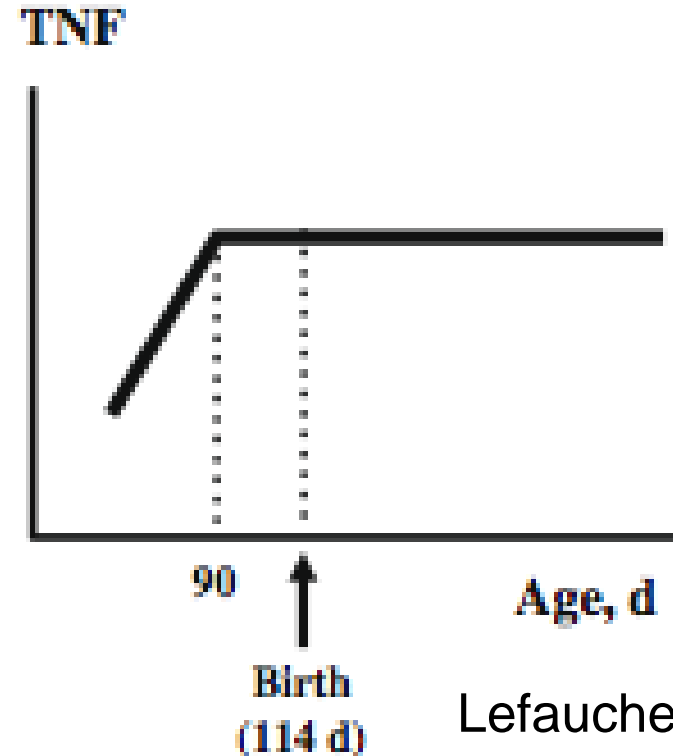
Review – Florini et al, 1996

# Skeletal muscle anatomy



## Muscle mass is related to.....

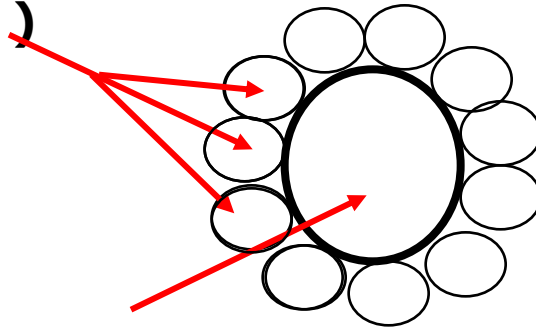
- Total number of fibres (TNF)
  - Hyperplastic growth
  - fixed at birth
- Cross Sectional Area (CSA) and Length
  - Hypertrophic growth



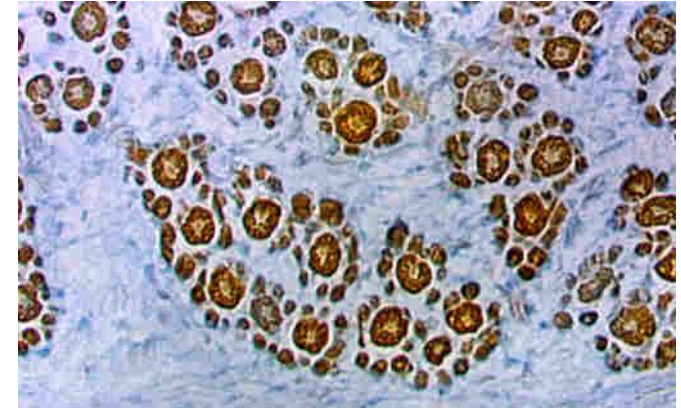
Lefaucheur 2010

# Primary and secondary fibre formation

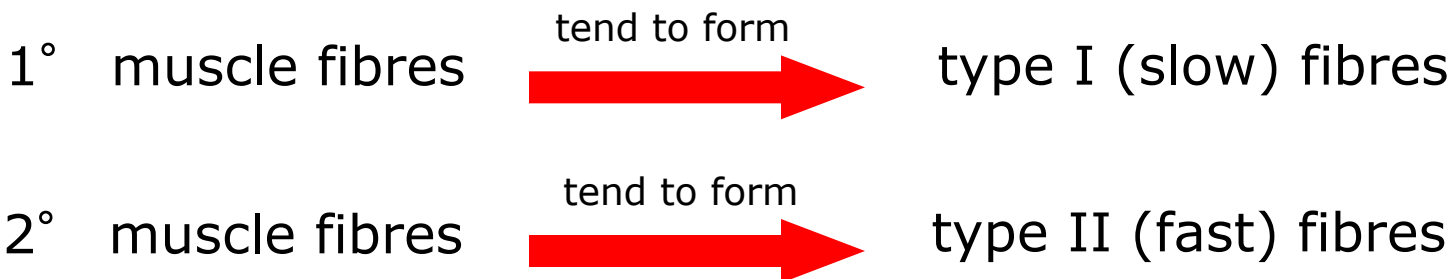
**Fast fibres (2° )**



**Slow fibre (1° )**



Pig muscle fibre formation  
(thanks to Dr C. Rehfeldt)



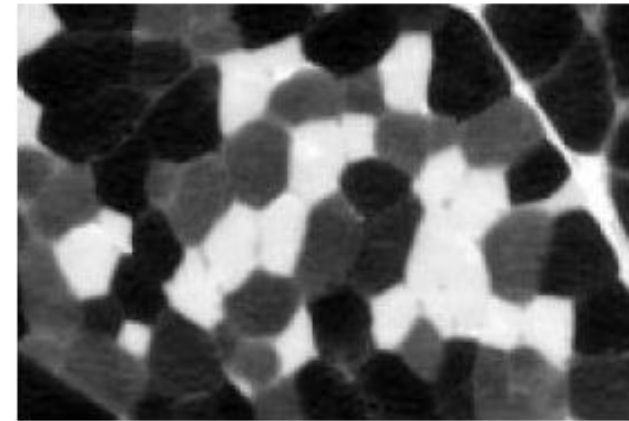
Proliferation and differentiation of myoblasts mainly takes place *in utero*



# Muscle fibre type

- Classic histochemistry staining
- Type I - slow oxidative (SO)
- Type IIA - fast oxidative glycolytic (FOG)
- Type IIB (or IIX) - fast glycolytic (FG)

Aberdeen Angus



Maltin et al 2001

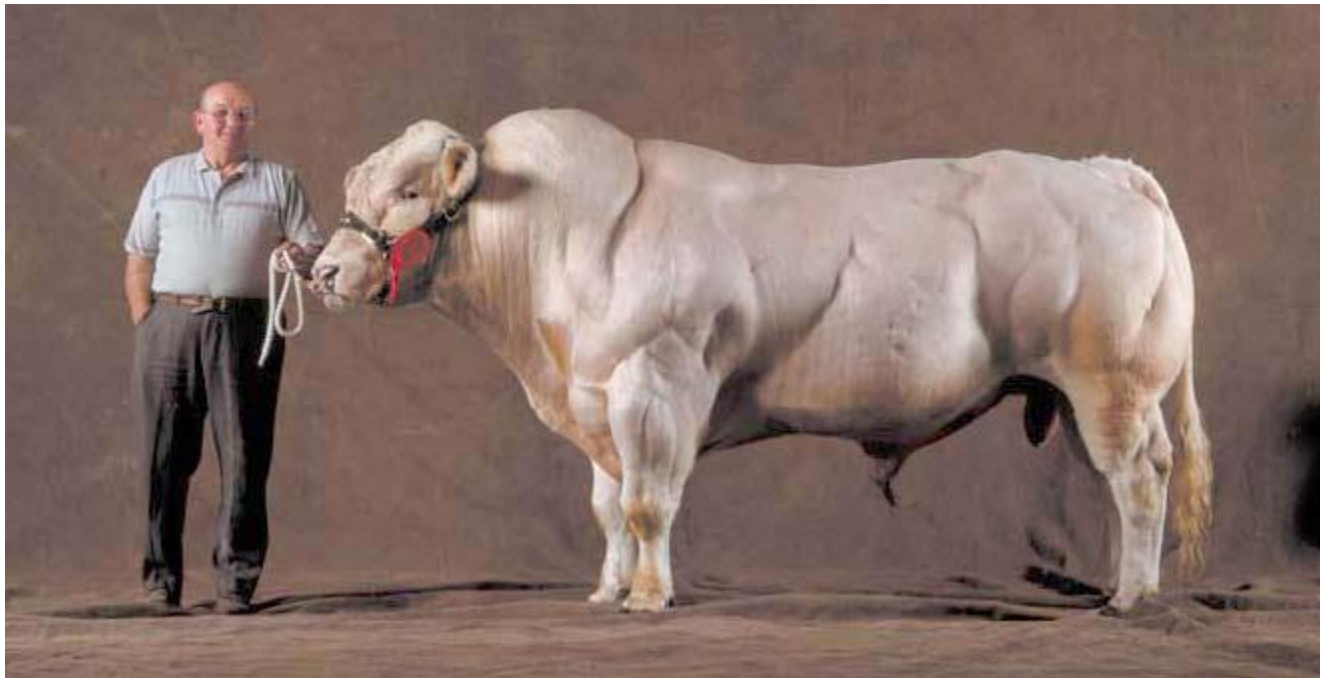
# Timings for appearance of fibres

	Primary	Secondary	Tertiary	Length of Gestation	Reference
<b>Poultry</b>	<b>3-7 df</b>	<b>8-16 df</b>	<b>-</b>	<b>21 days</b>	<b>Bandman &amp; Rosser (2000)</b>
<b>Rat</b>	<b>14-16 df</b>	<b>17-19 df</b>	<b>-</b>	<b>22 days</b>	<b>Wilson et al (1988)</b>
<b>Guinea pig</b>	<b>30 df</b>	<b>30-35 df</b>	<b>-</b>	<b>68 days</b>	<b>Dwyer et al (1995)</b>
<b>Pig</b>	<b>35 df</b>	<b>55 df</b>	<b>0-15 dpn</b>	<b>114 days</b>	<b>Lefaucheur et al (1995)</b>
<b>Sheep</b>	<b>32 df</b>	<b>38 df</b>	<b>62-76 df</b>	<b>145 days</b>	<b>Wilson et al (1992)</b>
<b>Bovine</b>	<b>60 df</b>	<b>90 df</b>	<b>110 df</b>	<b>278-283 days</b>	<b>Gagniere et al (1999)</b>
<b>Human</b>	<b>56 df</b>	<b>90 df</b>	<b>110-120 df</b>	<b>280 days</b>	<b>Draeger et al (1997)</b>

Adapted from Picard et al. (2002) and Brameld et al. (2003).

# Genetic effects on myogenesis

The greater TNF the greater the capacity for growth



double muscling

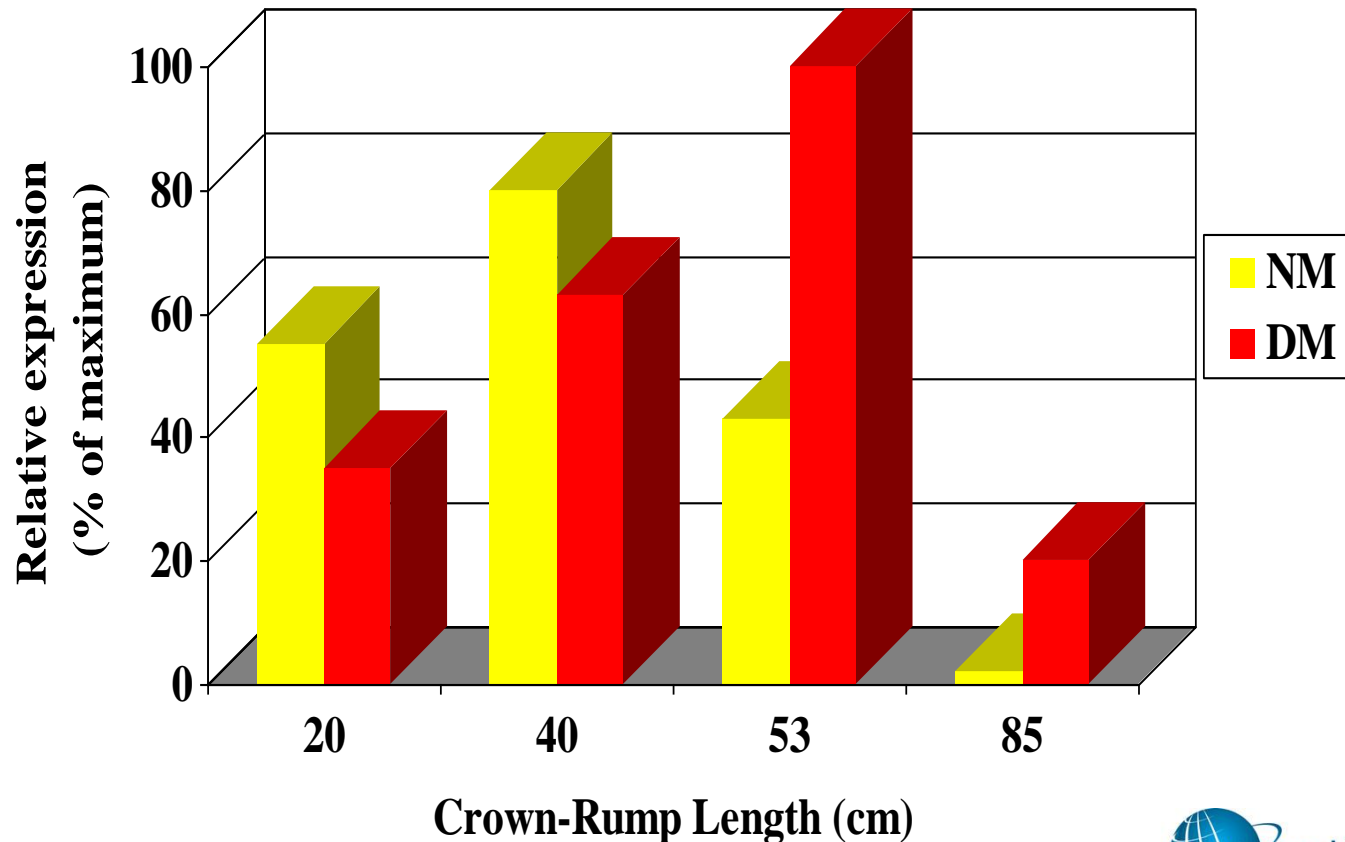


## Double muscling



- Increased TNF at birth
- Increased mitogenic activity of fetal blood (Gerrard & Judge, 1993)
- Mutation in myostatin gene (also called GDF-8, TGF $\beta$  family)
- Results in inactive protein being produced
  - TGF $\beta$  inhibits proliferation (and differentiation)  $\rightarrow$  removed an inhibitor
- More fibres due to more cell proliferation?
- BUT some cattle and pigs have similar mutations and no double muscling?
- Something else involved?

# IGF-II gene expression in developing fetal ST muscle of normal (NM) and double muscled (DM) cattle



Gerrard & Grant (1994)



## Double muscling



- Increased TNF at birth
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- Results in inactive protein being produced
  - TGF $\beta$  inhibits proliferation (and differentiation)  $\rightarrow$  removed an inhibitor.
- More fibres due to more cell proliferation?
- Delay in maximal IGF-II expression
- Delay in terminal differentiation?
- More fibres due to more cell proliferation AND delayed differentiation?

# Maternal nutrition & muscle fibre number

- Muscle fibre number in pigs, sheep and cattle is set at birth
- Therefore factors regulating this are active during gestation
- In pigs: runts are born with fewer muscle fibres and more intramuscular fat

## Comparisons between large and runt littermate groups

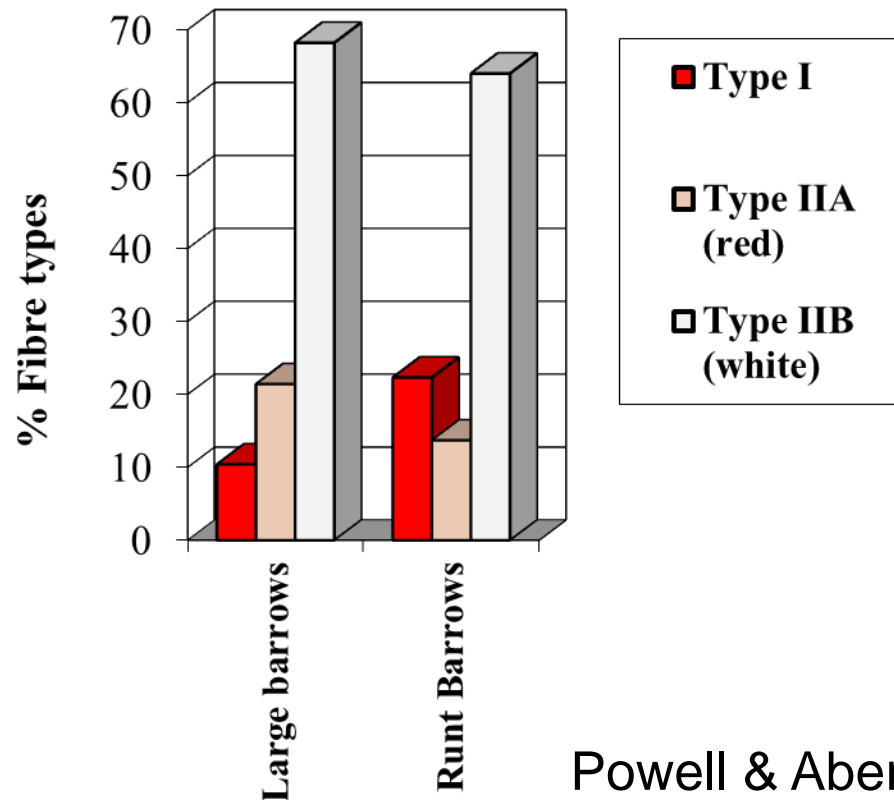


Parameter	Large piglet	Runt piglet
Birth Weight (g)	1544	776
ST 2°:1° fibre ratio	25.3	22.1
Trapezius 2°:1° fibre ratio	22.9	20.7

Handel & Stickland (1987)



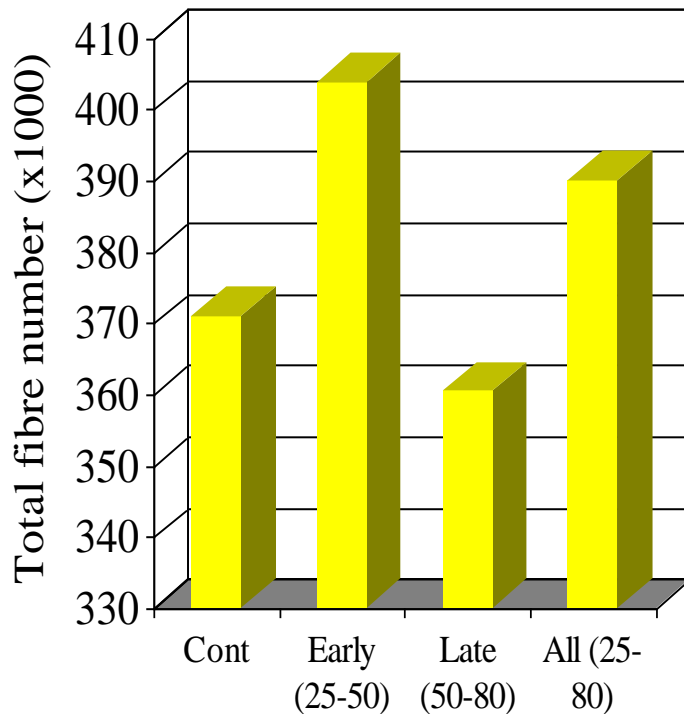
# SM muscle fibre types in runt and large birth weight pigs



- Runts have a higher proportion of type I fibres and fewer type II fibres than larger birth weight pigs

Powell & Aberle (1981)

# Effects of Maternal Nutrition in Pigs on ST TNF of Progeny at Birth



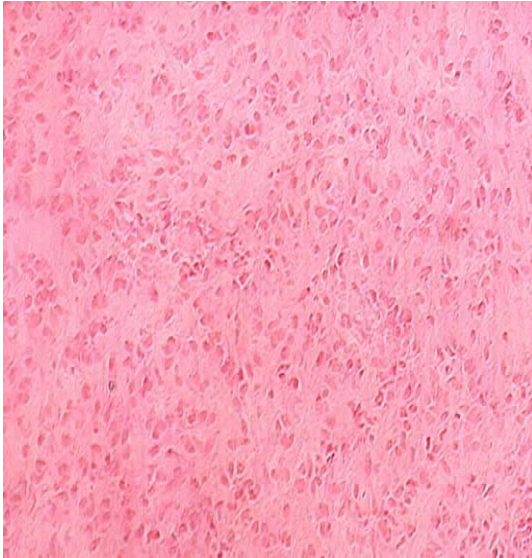
Dwyer et al (1994)

- Doubling maternal intake at early gestation (25-50 days) increased ST TNF
  - Mainly in smaller littermates
  - Normal practice is under-nutrition
- Increase in no. of secondary fibres, no effect on primary fibres
- May eliminate runts
  - no effect on large littermates
  - already reaching genetic potential ?

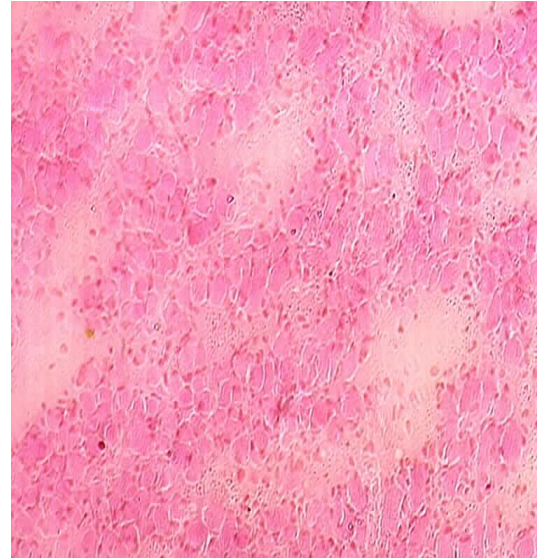
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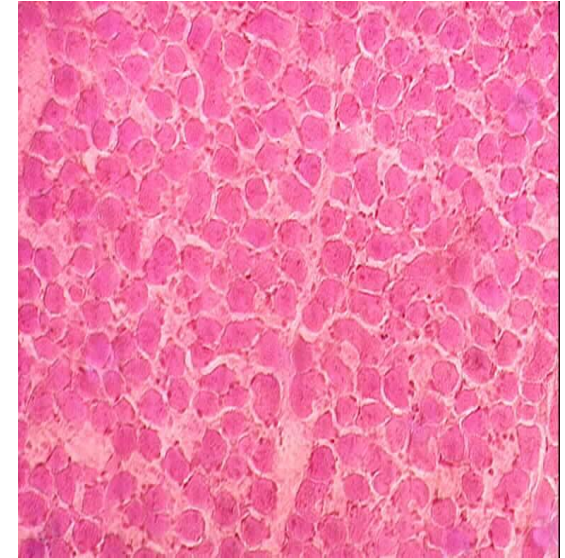
# Fibre formation in fetal sheep



**d70**



**d85**



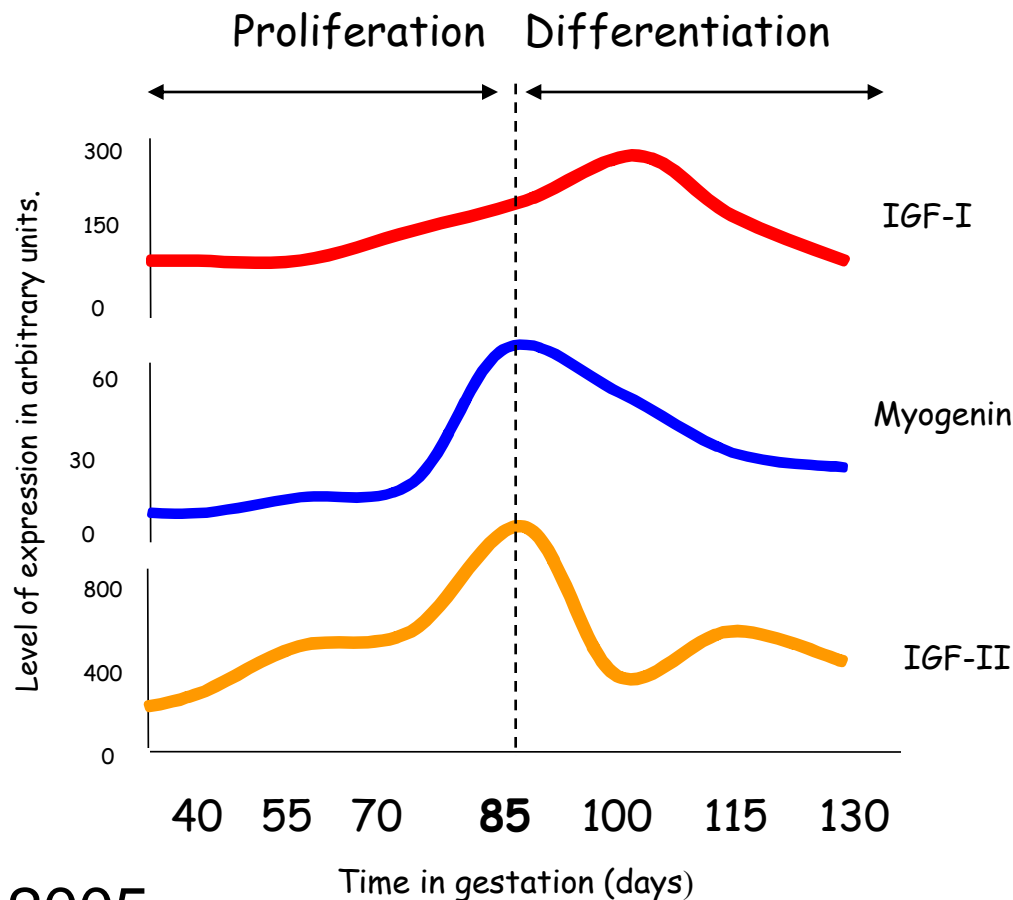
**d100**

**H&E stained muscle sections**



Fahey et al, 2005

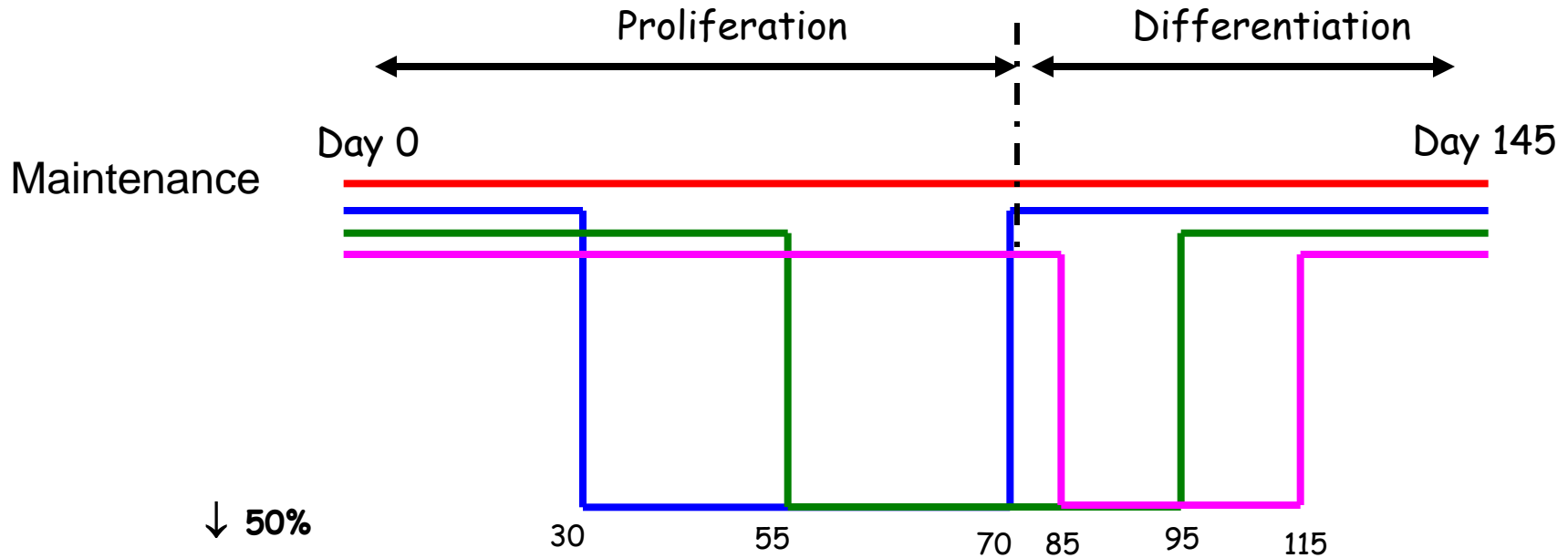
# Ontogeny of factors thought to control muscle development (sheep)



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# Ewe feeding trial - Fahey et al, 2005



**d30-d70 during proliferation**

**d55-d95 during both proliferation and differentiation**

**d85-d115 during differentiation**

32 ewes, 8 per group



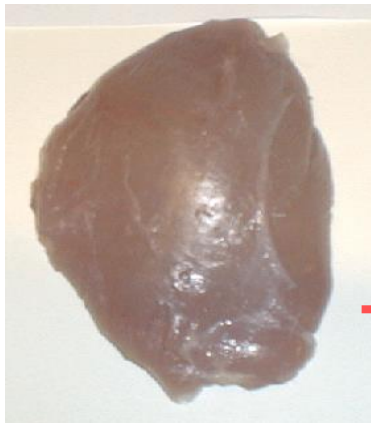
# Location of muscles



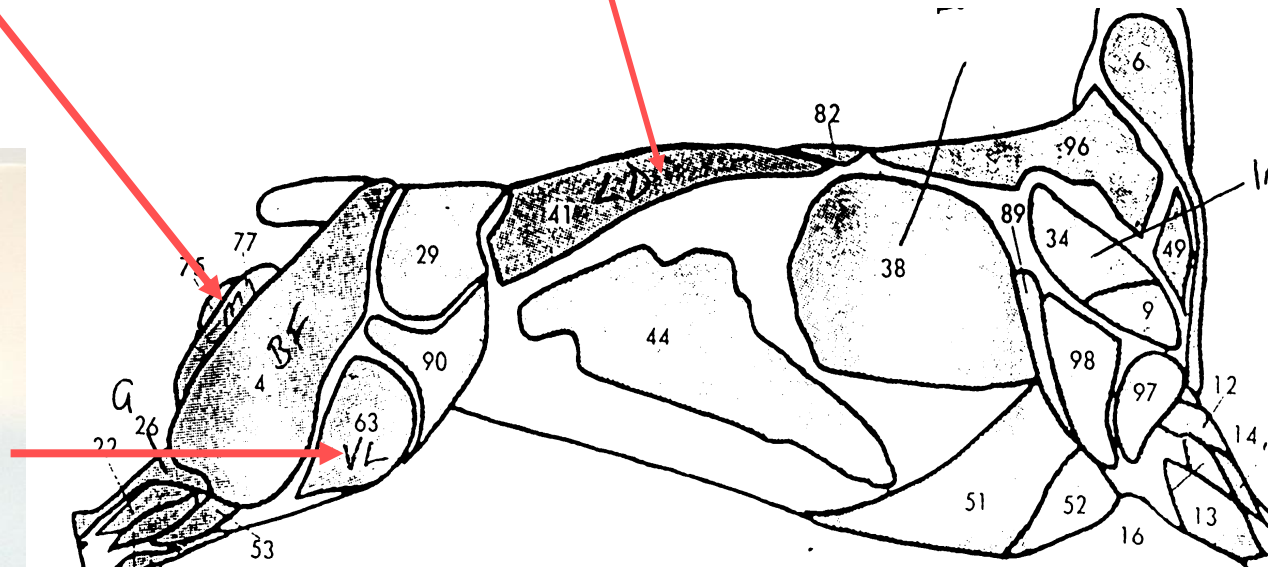
ST



LD

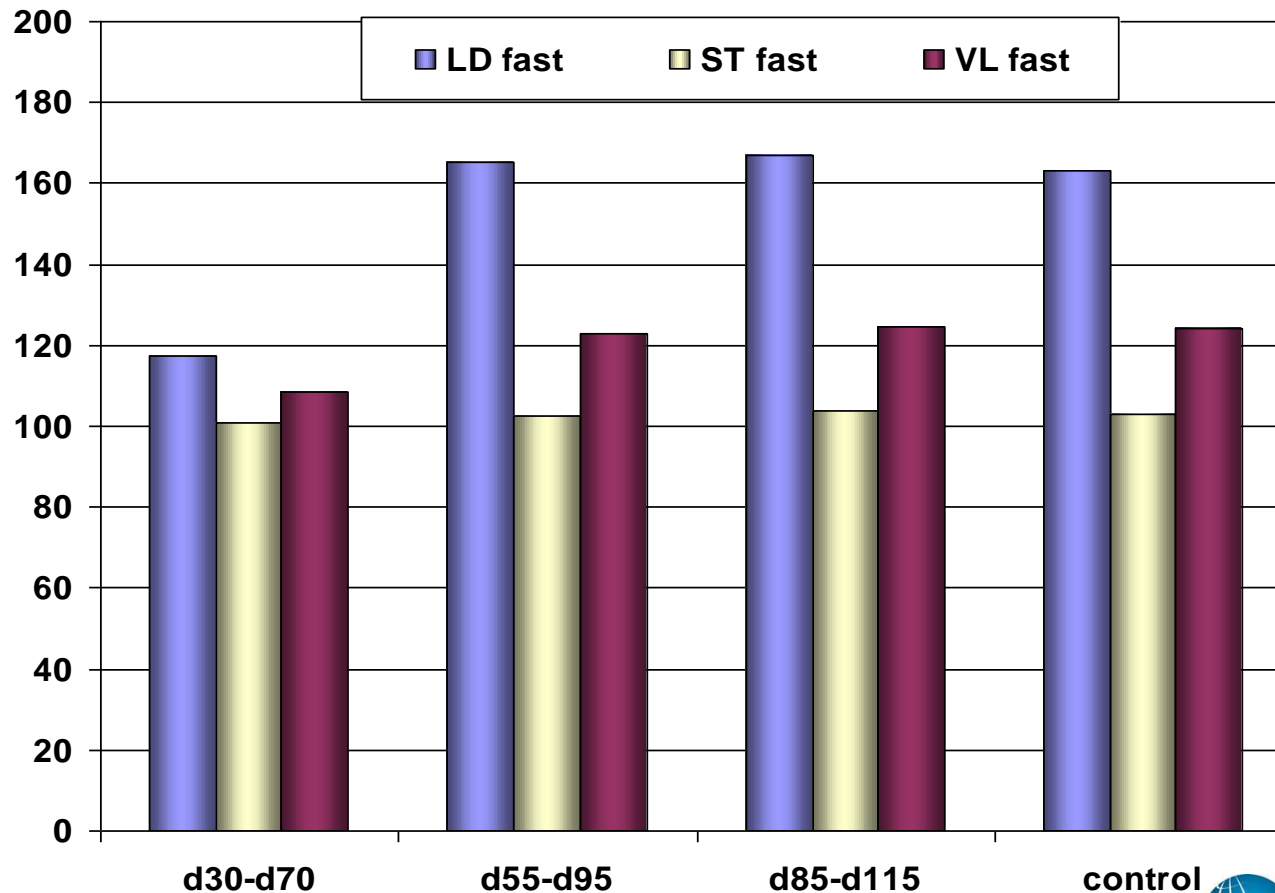


VL





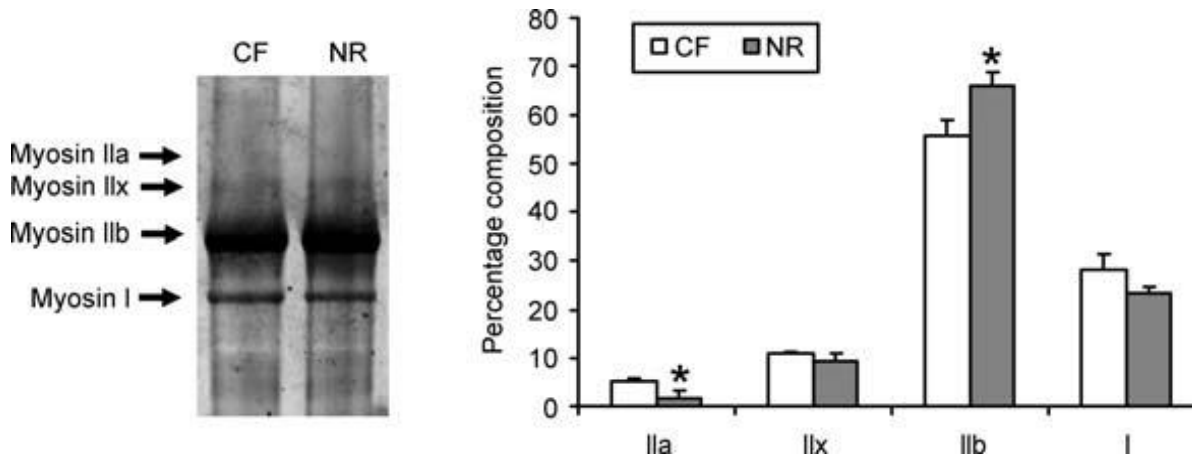
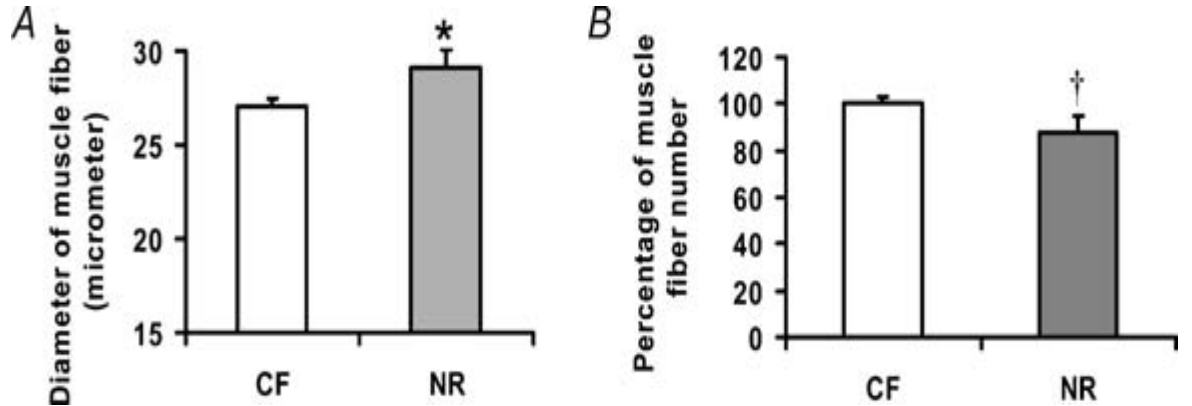
# Numbers of fast fibres (14d)



Fahey et al, 2005b

# Opposite effects seen in older lambs (Zhu et al, 2006)

Restricted (50 v 100%) d28-78, male lambs @ 8months (34-35wks), LD muscle taken



We observed very similar results at younger ages (17 and 24 weeks - Daniel et al 2007)

# Nutritional effects on TNF

- Effects of maternal nutrition on numbers of muscle fibres in offspring observed in various species
  - Guinea-pigs, rats, pigs, sheep

• Low(er) nutrition   no. of 2° fibres

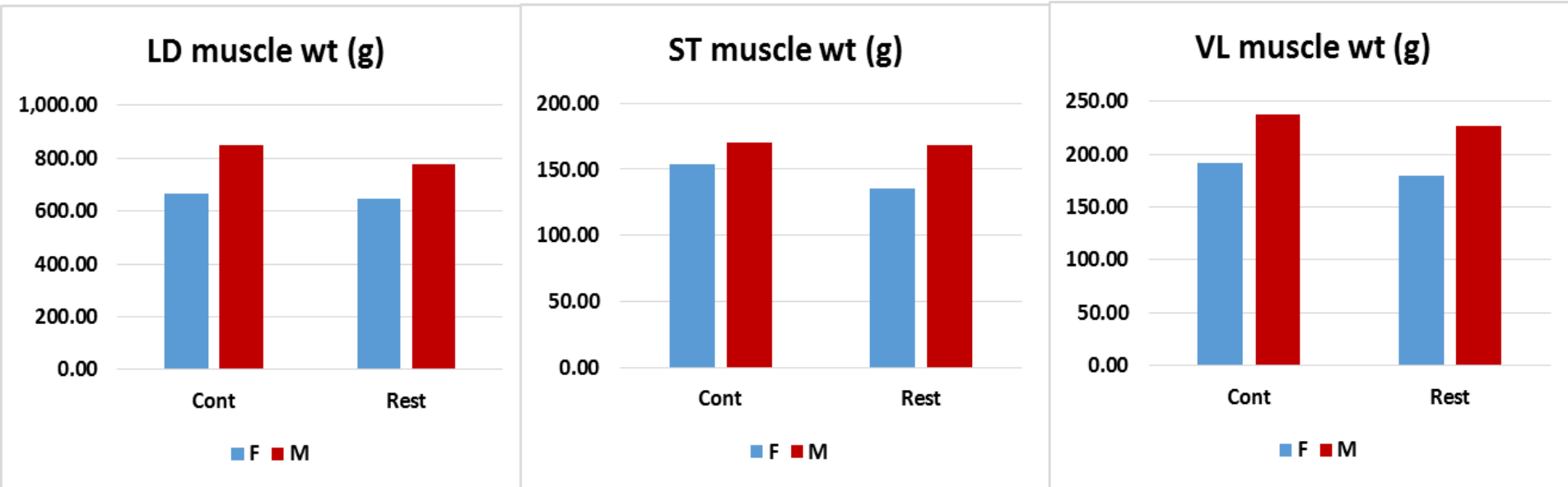
• High(er) nutrition   no. of 2° fibres

- Timing of intervention is **CRITICAL**
- Normally just **BEFORE** fibre formation
- Long-term effects are unclear!!!

# Effects on muscle wts in older lambs (Daniel et al, 2007)



Restricted (50 v 100%) d30-85, male & female lambs @ 24wks



$F < M; \text{Rest} < \text{Cont}$

$F < M$

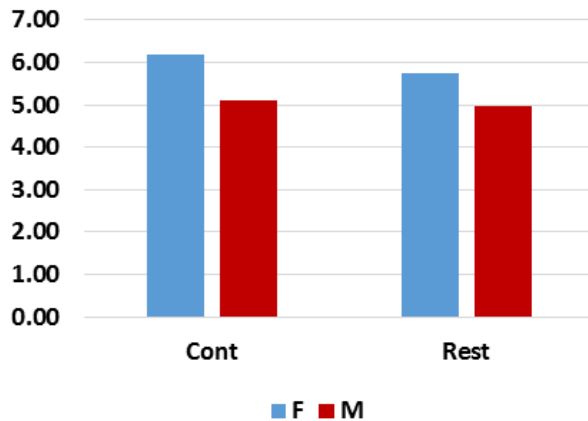
$F < M; \text{Rest} < \text{Cont}$

# Effects on adiposity in older lambs (Daniel et al, 2007)



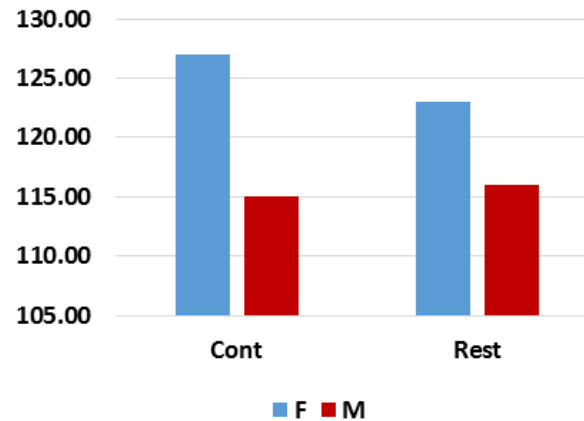
Restricted (50 v 100%) d30-85, male & female lambs @ 24wks

**Back fat thickness (mm)**



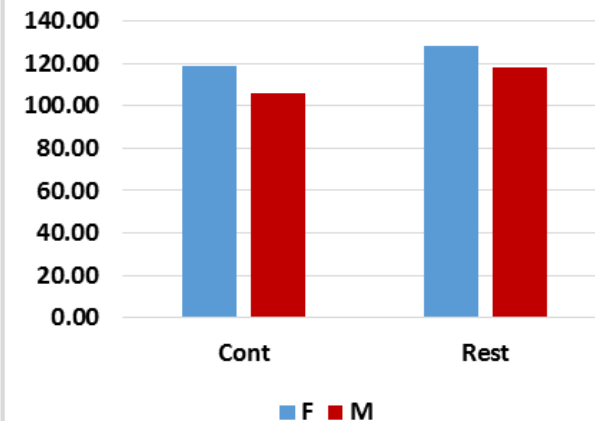
F>M

**Omental adip diam**



F>M

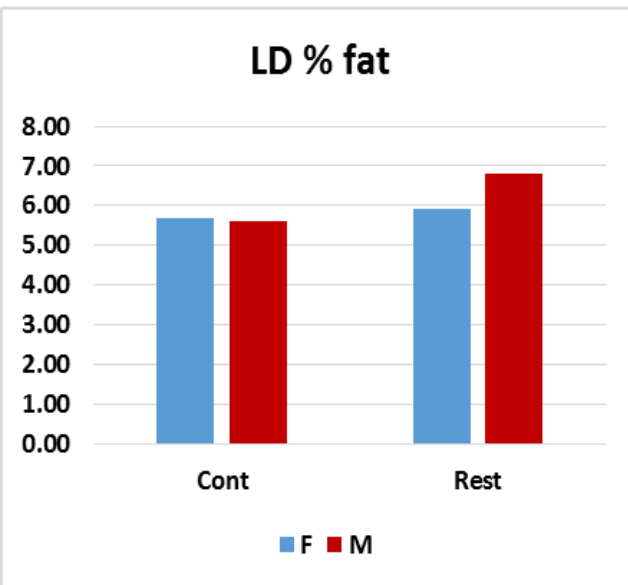
**Perirenal adip diam**



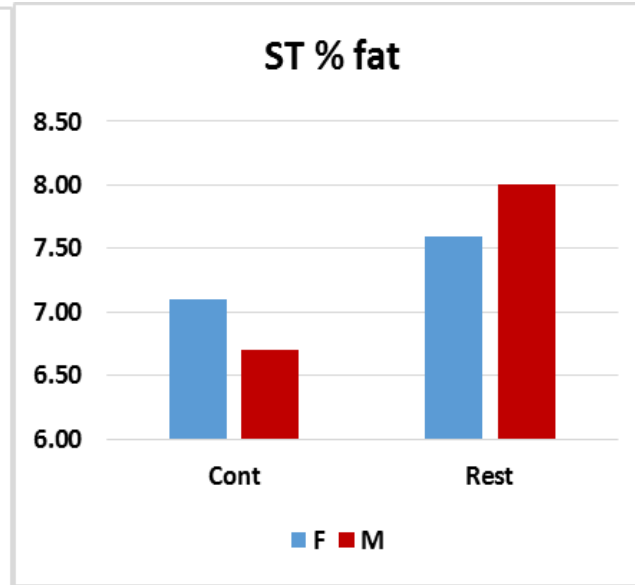
F>M; Rest>Cont

# Effects on muscle % fat in older lambs (Daniel et al, 2007)

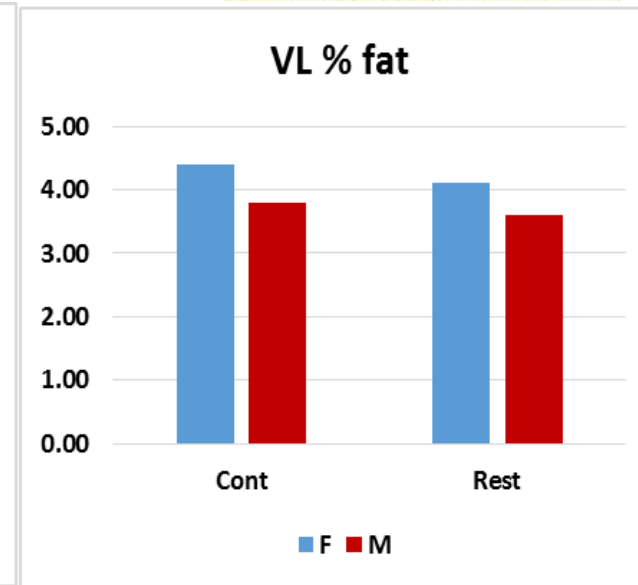
Restricted (50 v 100%) d30-85, male & female lambs @ 24wks



RestM > ContM



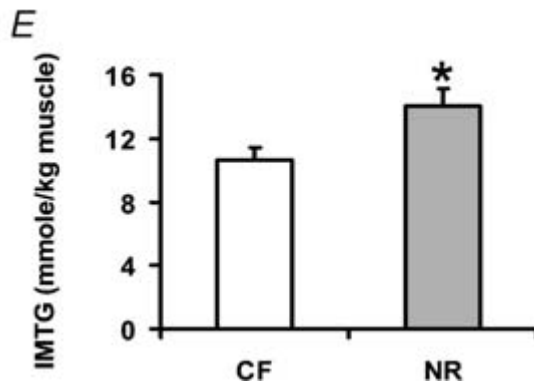
Rest > Cont



F > M

# Effects on adiposity in older lambs (Zhu et al, 2006)

Restricted (50 v 100%) d28-78, male lambs @ 8months (34-35wks), LD muscle taken







## Zhu et al, 2006

Restricted (50 v 100%) d28-78, male lambs 8months old (34-35wks)

Category	CF	NR
Liveweight (kg)	56.80 ± 1.23	61.72 ± 1.62*
Carcass wt (kg)	28.83 ± 0.92	31.62 ± 1.01 <sup>†</sup>
Kidney + pelvic fat (kg)	0.46 ± 0.05	0.68 ± 0.06*
LD muscle (g)	776.5 ± 12.1	775.7 ± 30.0
ST muscle (g)	166.4 ± 4.4	169.3 ± 3.9
K + P fat (%CW)	1.66 ± 0.12	2.18 ± 0.20*
LD muscle (%CW)	2.71 ± 0.11	2.46 ± 0.08 <sup>†</sup>
ST muscle (%CW)	0.58 ± 0.02	0.54 ± 0.02 <sup>†</sup>



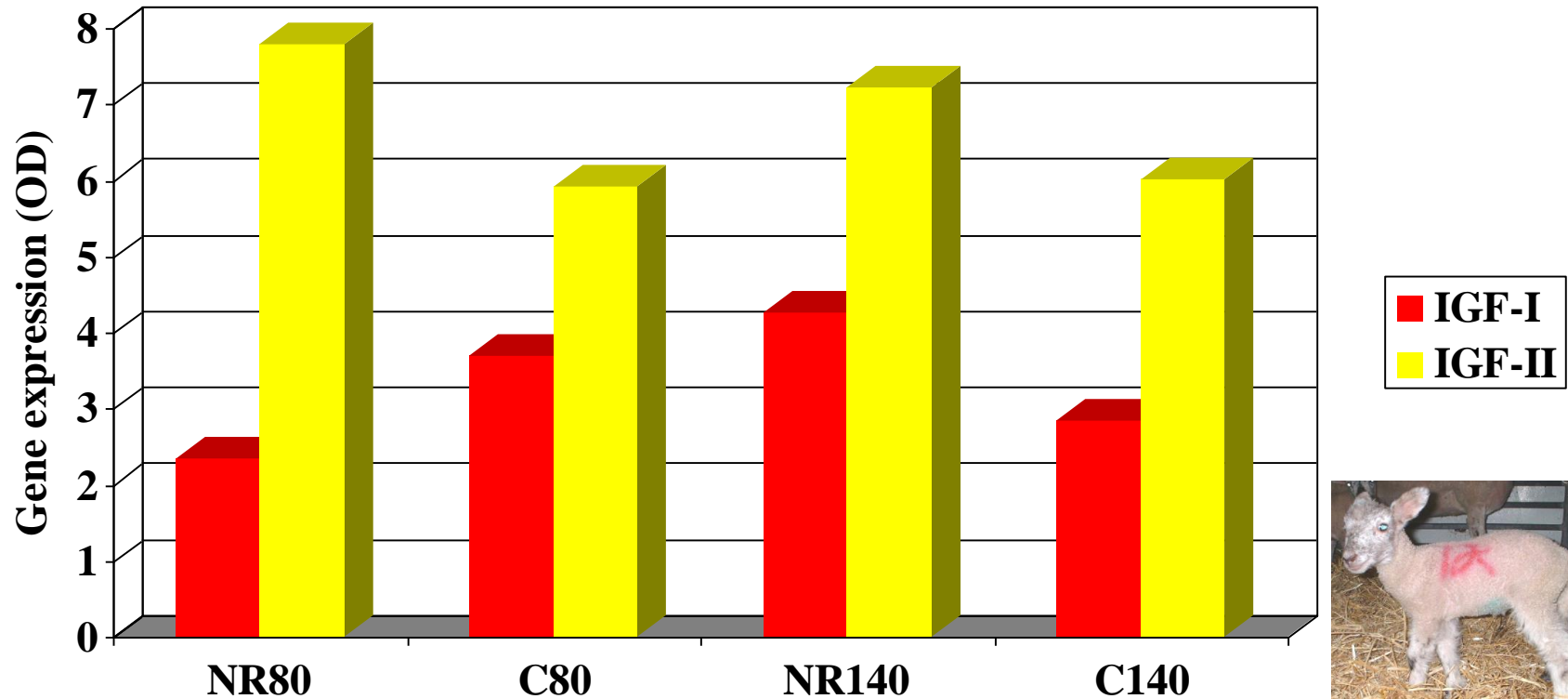
# Nutritional effects on body composition

- Evidence that there may be long-term effects on muscle weights and some measures of adiposity
- Low(er) nutrition   muscle wts
- Low(er) nutrition   adiposity
- Dependent upon the age of the lamb
  - Increased adiposity seen at 24 and 34-35wks, but not 17wks

## Possible Mechanism

- Nutrition regulates the IGF axis
- High nutrition increases hepatic IGF-I production (and plasma IGF-I)
  - Does high(er) plasma IGF-I stimulate myoblast proliferation and inhibit muscle IGF-II expression and differentiation?
- Does low nutrition have opposite effect (reduced plasma IGF-I, reduced proliferation, increased IGF-II and early differentiation)?
- Is muscle IGF-II expression the KEY regulator of differentiation and fibre formation?

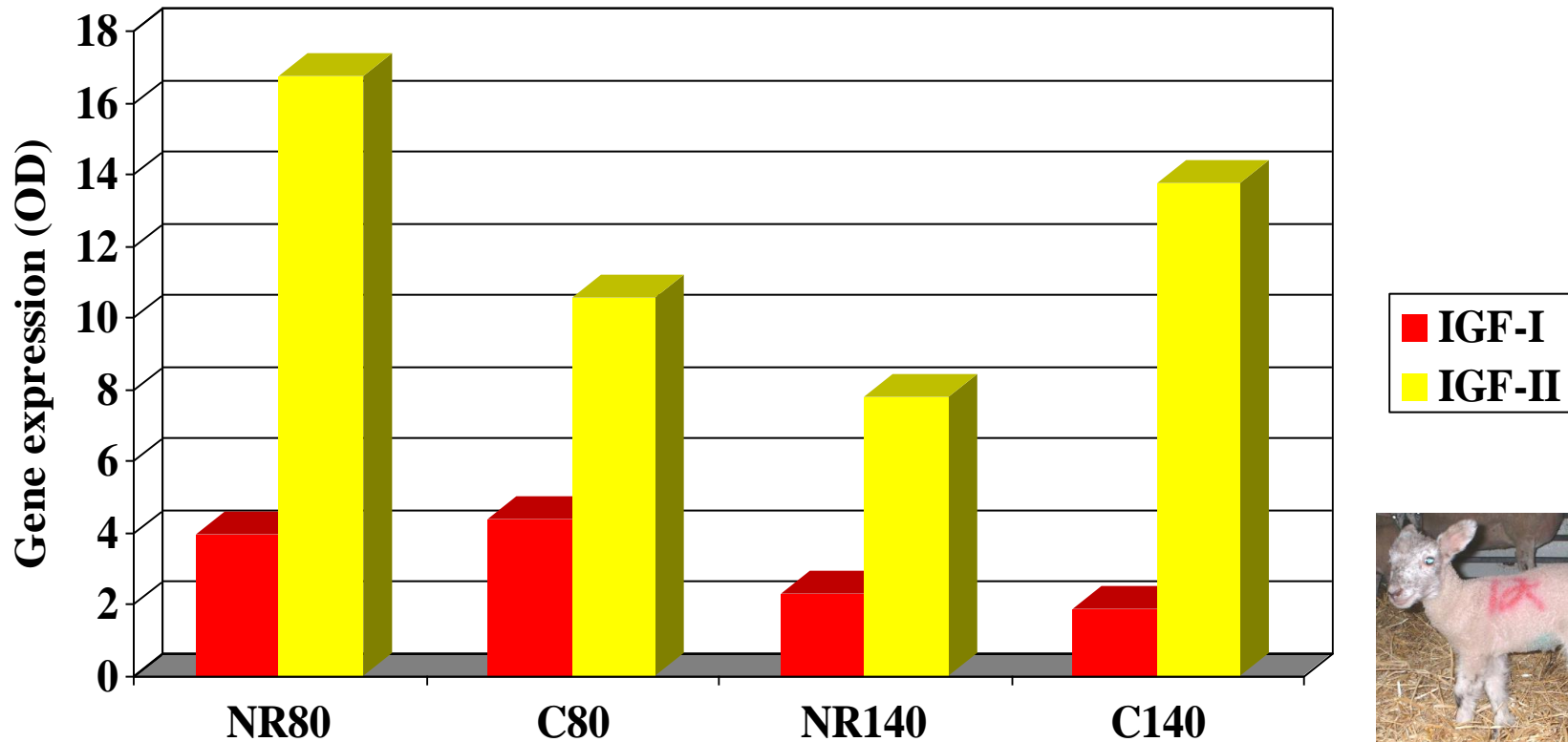
# Effects of maternal nutrition on fetal sheep liver gene expression (Brameld et al, 2000)



C = 150%  
NR = 60%

	Age	Diet	Age x Diet
IGF-I	NS	NS	0.061
IGF-II	NS	0.061	NS

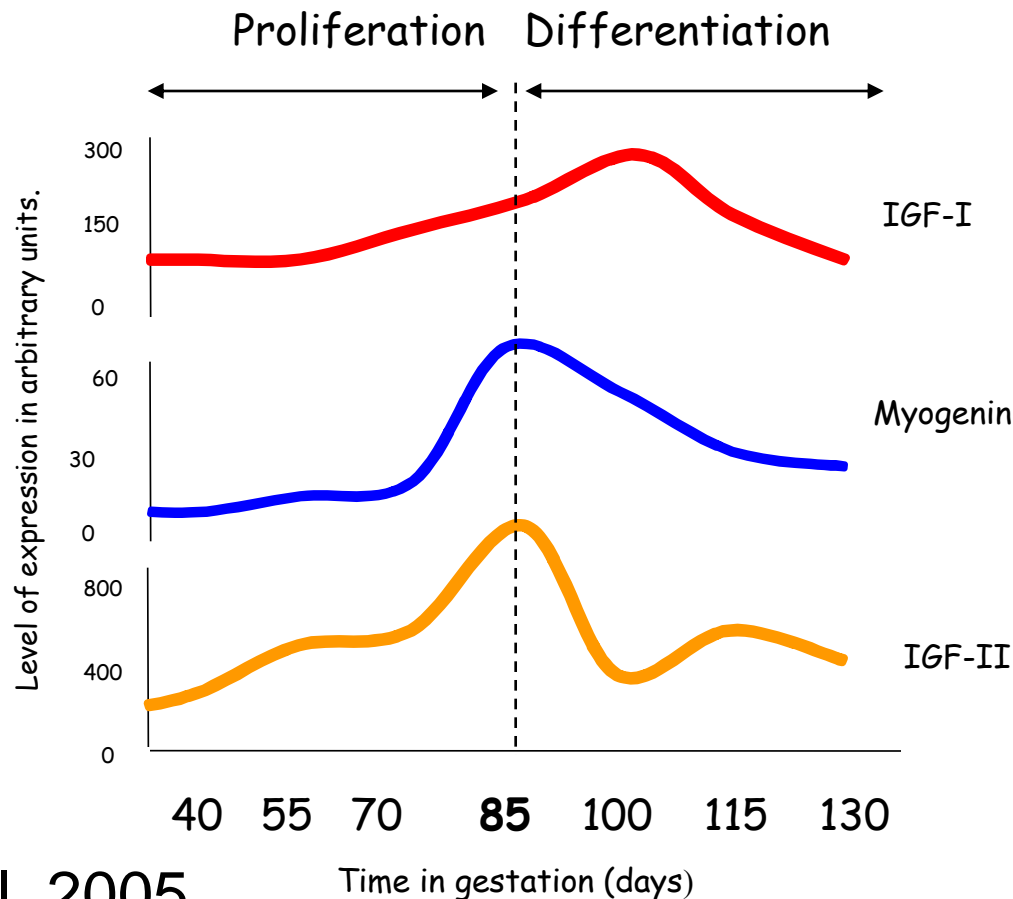
# Effects of maternal nutrition on fetal sheep muscle gene expression (Brameld et al, 2000)



C = 150%  
NR = 60%

	Age	Diet	Age x Diet
IGF-I	<0.001	NS	NS
IGF-II	0.056	NS	<0.001

# Ontogeny of factors thought to control muscle development (sheep)



## Conclusions: Not all fibres are affected

- The effects of environment/nutrition are mainly on secondary fibres formed
  - no. of primary fibres genetically determined ?
  - Critical window prior to major fibre formation (species differences!!!)
  - Very few studies have grown offspring to older ages,
    - but effects may be lost/ compensated for by good postnatal nutrition (more work needed)
  - BUT TFN is difficult to measure in large muscles!!!
  - Long-term effects on muscle wts and adiposity
  - Is muscle IGF-II a key factor relating to timing of differentiation/ fibre formation?

# Acknowledgements

## Collaborators:

Peter Buttery

Jim Craigon

Nigel Scollan (IBERS)

## Funding:



## Researchers & Technical Support:

Angela Fahey

Deb Surgay

John Corbett

Chris Essex

Lesley Stubbings